

Fatty Acid Composition and Mineral Contents of Pea Genotype Seeds

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ABSTRACT: Metal, non-metal and heavy metal contents of different pea genotype seeds were determined by Inductively Coupled Plasma Atomic Emission Spectrometry (ICP-AES). For all genotypes, significant differences were observed in the mineral contents. Potassium was the most abundant element, ranged from 10146.13 mg/kg (PS3048) to 13171.97 mg/kg (PS3053) (Table 1). In addition, the phosphor content of pea seeds was found between 4004.31 mg/kg (PS 30100) and 5651.27 mg/kg (PS 3057). These pea genotypes contained 1562.32 mg/kg to 2034.28 mg/kg magnesium. Zinc contents of pea samples changed between 29.66 mg/kg (PS 3055) and 67.81 mg/kg (PS 4053 B). The oil contents of pea samples ranged from 0.84% (PS4053 B) to 3.59% (PS 3055). Oleic acid is predominant fatty acid 12.95% to 45.02% followed by palmitic 13.68% to 77.28%, stearic (1.66% to 15.99%) acids. The highest oleic acid was found in PS3048 genotype (45.02%). The highest palmitic acid was found in PS4021 pea sample (77.28%). The current study contributes to the available information concerning the composition of several pea genotypes grown in Turkey.

KEYWORDS: Pea; Genotypes; Oil; Protein; Mineral; Fatty acid composition; GC; ICP-AES.

INTRODUCTION

Pea (*Pisum sativum* L.) is one of the important legumes grown in the world. It supplies adequate amounts of protein and minerals in the diet [1-6]. Progress in pea breeding results not only in higher yields but also in changes in the chemical composition of seeds [7,8]. Legume seeds are rich in mineral elements [9]. Nutrition to the seed crop may improve seed quality [10].

The oil content of pea seeds ranged from 0.8 to 6.1% [11]. In addition, Coxon and Davies [12] reported that wrinkled and round seeded peas contained 4.5-5.2% and 2.8-3.1% crude lipid, respectively. Although low oil, it may be of importance in the flavour of peas [13]. Pea is an annual self-pollinated species, and highly valued food legume grown extensively in the world. In Turkey,

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the pea is considered to be a health vegetarian food and it is one of the most important human nutrition [14]. The aim of the current study was to investigate the fatty acid composition and mineral contents of some pea genotypes grown in Konya in Turkey.

EXPERIMENTAL SECTION

Material

In this study, PS30100, PS3029, PS3053, PS4021, PS4053 B, PS3048, PS3055, PS3057, and PS4028 pea genotypes that shown differences for morphological and high yield were used.

Methods

After about 50 g pea sample was dried at 65 °C for 24 h, they were ground with a hummer mill. Protein contents were determined by the Kjeldahl apparatus. 6.25 was used as a nitrogen coefficient. The crude protein content of the grains was calculated by the coefficient of nitrogen quantity (6.25) [15].

Oil extraction

About 2 g of the seeds were ground in a ball mill and extracted with petroleum ether in a Soxhlet apparatus for 6 h [16]. The solvent was removed by a rotary evaporator at 40 °C and 25 mmHg. The oil was dried by a stream of nitrogen and stored at – 20 °C until used.

Determination of fatty acids

Fatty acid compositions for pea seed oil were determined using a fatty acid methyl ester method as described by *Hışıl* [17]. The oil was extracted three times for 2 g air-dried seed sample by homogenization with petroleum ether. The oil samples (50-100 mg) was converted to its Fatty Acid Methyl Esters (FAME). The methyl esters of the fatty acids (1 µL) were analysed in gas chromatography (GC-MS (Gas Chromatography-Mass Spectrometry) Agilent Technologies HP 6890) and MS; Agilent Technologies 5975C VL MSD equipped with a Flame Ionising Detector (FID), a fused silica capillary column (60 m x 0.25 mm i.d.; film thickness 0.250 micrometer). It was operated under the following conditions: oven temperature program. 175 °C for 7 min. Raised to 250 °C at a rate of 5 °C/min and then kept at 250 °C for 15 min; injector and detector temperatures, 250 and 250 °C; respectively, carrier gas. Nitrogen at flow rate of 1.51 mL/min; split ratio. 1/50 µL/min.

Determination of mineral contents

Pea genotype samples were dried at 70 °C in a drying cabinet with air-circulation until they reached constant weight. Later, about 0.5 g dried and ground samples were digested by using 5mL of 65% HNO₃ and 2 ml of 35% H₂O₂ in a closed microwave system (Cem-MARS Xpress). The volumes of the digested samples were completed to 20 mL with ultra-deionized water, and mineral contents were determined by Inductively Coupled Plasma Atomic Emission Spectrometry (ICP AES) (Varian-Vista, Australia). Measurements of mineral concentrations were checked using the certified values of related minerals in the reference samples received from the National Institute of Standards and Technology (NIST; Gaithersburg, MD, USA) [18].

Working conditions of ICP-AES

Instrument	: ICP-AES (Varian-Vista)
RF Power	: 0.7-1.5 kW (1.2-1.3 kw for Axial)
Plasma gas flow rate (Ar):	10.5-15 L/min. (radial) 15 (Axial)
Auxiliary gas flow rate (Ar)	: 1.5
Viewing height	: 5-12 mm
Copy and reading time	: 1-5 s (max.60 s)
Copy time	: 3 s (max. 100 s)

Statistical analysis

Experiment "randomized block" established in three replicates patterns in the experiment field of Faculty of Agriculture, Selçuk University. Varyans analysis and LSD test was made using "MSTAT-C" packet program access on the computer [19].

RESULTS AND DISCUSSION

The mineral contents of pea genotypes are given in Table 1. There were significant differences in these values among the nine genotypes of a pea. These differences in mineral and oil contents were due to a combination of genetic and environmental factors. For all genotypes, significant differences were observed in the mineral contents. Potassium was the most abundant element, ranged from 10146.13 mg/kg (PS3048) to 13171.97 mg/kg (PS3053) (Table 1). In addition, the phosphor content of pea seeds was found between 4004.31 mg/kg (PS 30100) and 5651.27 mg/kg (PS 3057). These pea genotypes contained 1562.32 mg/kg to 2034.28 mg/kg magnesium. Zinc contents of pea samples

Table 1: Protein (%) and mineral contents of pea genotype seed (mg/Kg).

Pea Genotypes	Protein	Al	Mo	Ca	B	Cd	Cu	Fe
PS30100	24.32 b	24.62 a	2.41 d	374.34 de	9.80 a	0.143	8.26 b	62.97 e
PS3029	21.52 de	23.45 ab	2.17 d	392.28 de	9.95 a	0.137	8.47 b	75.91 ab
PS3053	22.96 c	19.33 cd	3.20 c	809.03 b	10.61 a	0.167	6.55 d	70.13 cd
PS4021	24.63 b	16.42 e	2.42 d	1179.99 a	10.45 a	0.147	7.28 cd	65.52 de
PS4053 B	23.99 b	21.90 b	3.25 c	1262.82 a	10.47 a	0.123	7.89bc	78.15 a
PS3048	22.28 cd	15.70 e	1.34 e	484.13 cd	10.44 a	0.167	10.13 a	78.69 a
PS3055	20.66 e	17.55 de	6.10 a	330.48 e	7.52 b	0.137	7.76bc	64.63 e
PS3057	24.00 b	21.45bc	4.96 b	447.74cde	7.86 b	0.140	8.15bc	71.45bc
PS4028	27.35 a	17.14 de	3.57 c	517.00 c	10.51 a	0.123	8.05bc	65.77 de
Mean	23.52	19.73	3.27	644.20	9.73	0.143	8.06	70.36
Lsd	1.02	2.48	0.46	117.3	1.75		0.96	5.43
Pea Genotypes	K	Mg	Mn	Na	Ni	P	S	Zn
PS30100	11640.86b	1663.91 de	13.47 cd	169.59bc	2.02 d	4004.31 c	2389.11abc	51.74bc
PS3029	11564.75b	1772.24bcd	13.38 cd	146.53 de	2.54bc	4049.06 c	2321.23 a-d	56.95 b
PS3053	13171.97 a	1909.70 ab	16.35 b	192.16 a	2.44 cd	4294.75 c	2447.92 ab	50.57 c
PS4021	11863.94b	1994.11 a	12.00ef	158.53 cd	4.67 a	5156.98 b	2464.00 a	42.37 d
PS4053 B	12343.30ab	2034.28 a	20.67 a	182.05 ab	2.14 cd	4287.58 c	2434.49 ab	67.81 a
PS3048	10146.13c	1562.32 e	10.83 g	157.48cde	2.46 cd	4157.50 c	2350.13abc	67.63 a
PS3055	12025.55b	1718.08 cd	12.78 de	142.89 e	2.23 cd	5064.63 b	2125.79 d	29.66 e
PS3057	13137.69 a	1829.13bc	14.35 c	147.35 de	2.28 cd	5651.27 a	2181.45 cd	33.41 e
PS4028	12208.34ab	1672.24 de	11.14fg	183.72 ab	2.93 b	5374.09 ab	2230.27bcd	33.25 e
Mean	12011.39	1795.11	13.89	164.48	2.64	4671.13	2327.16	48.16
Lsd	963.50	146.70	1.10	15.34	0.45	430.60	223.00	5.225

changed between 29.66 mg/kg (PS 3055) and 67.81 mg/kg (PS 4053 B). Harmankaya et al. [20] reported that several pea genotypes contained 45.91-157.4 mg/100g Ca, 47.31-102.8 mg/100g Mg, 562.8 mg/100g-937.8 mg/100 g K, and 163.4-374.2 mg/100 g P and 2.10-5.71 mg/100g Zn. Wang and Daun [21] reported that Canadian field peas contained 59.6-106.9 mg/100g Ca, 4.1-7.9 mg/100g Fe, 687.4-1473.2 mg/100g K, 115.4-172.3 mg/100g Mg, 226.5-950.5 mg/100g P and 2.5-6.4 mg/100g Zn. The results are partly similar to the finding of Wang and Daun [21] and Harmankaya et al. [20].

The oil and fatty acid composition of pea genotype seed oils are presented in Table 2. The oil contents of pea samples ranged from 0.84 (PS4053 B) to 3.59% (PS 3055).

In the previous study, pea seed contained 0.24 to 4.97% oil [22-24]. Oleic acid is predominant fatty acid 12.95% to 45.02% followed by palmitic 13.68 to 77.28%, stearic (1.66 to 15.99%) acids. The highest oleic acid was found in PS3048 genotype (45.02%). The highest palmitic acid was found in PS4021 pea sample (77.28%). Wang and Daun [21] reported that Canadian field pea oils contained 0.30-0.67% myristic, 8.57-12.73% palmitic, 2.39-5.27% stearic, 17.83-30.43% oleic, 40.55-52.44% linoleic, 10.25-17.15% linolenic, 0.50-1.19% arachidic acids. Ryan et al. [25] reported that pea oil contained 10.65% palmitic, 3.29% stearic, 28.25% oleic, 47.59% linoleic, 9.29% linolenic and 0.22% arachidic acids. Srivastava et al. [26] reported that field pea oil contained 12.0-18.4% palmitic,

Table 2: Oil contents and fatty acid composition of pea seed oil (%).

Genotypes	Oil	Myristic C14:0	Palmitic C16:0	Stearic C18:0	Oleic C18:1	Linoleic C18:2	Linolenic C18:3	Arachidic C20:0	Eicosenoic C20:1
PS30100	1.439 d	2.257a	32.877e	15.990 a	44.800 b	0.000 g	0.000 f	0.000 g	0.000 e
PS3029	1.761 cd	0.807f	33.173 d	6.349 d	44.573 c	6.903 f	4.587 c	2.077 e	1.346 c
PS3053	1.420d	0.656 h	13.536 i	1.660 i	21.933 g	51.335 a	10.882 a	0.000 g	0.000 e
PS4021	2.082bc	0.930 e	77.275 a	4.053 f	0.436 i	13.883 d	0.000 f	0.000 g	0.000 e
PS4053 B	0.835e	1.911b	30.146 g	3.236 g	42.922 d	0.000 g	0.000 f	4.581 a	1.324 d
PS3048	2.406b	1.255d	30.479 f	6.793 c	45.024 a	9.585 e	1.055 e	2.174 d	0.000 e
PS3055	3.594a	0.778g	40.629 c	6.316 e	26.032 f	14.803 c	3.154 d	3.027 c	3.101 a
PS3057	1.583cd	0.393 i	13.678 h	1.850 h	39.634 e	36.792 b	7.017 b	0.640 f	0.000 e
PS4028	1.320de	1.781 c	54.584 b	10.652 b	12.950 h	0.000 g	0.000 f	3.546 b	1.474 b
Mean	1.827	1.196	36.264	6.322	30.922	14.811	2.966	1.783	0.805
Lsd	0.510	0.005	0.030	0.010	0.006	0.003	0.007	0.005	0.002

2.0-4.2% stearic, 16.5-24.1% oleic, 37.9-53.9% linoleic and 6.8-10.6% linolenic acids. Pea seed oil contained 18.64% palmitic, 50.72% linoleic, 11.3% linolenic, 5.27% oleic and 4.1% stearic acids [27]. This is oil content and its variation during seed growth is a function of variety [28]. Welch and Griffiths [22] reported that pea oil contained 6.4-13.4% linolenic, 43.7-60.9% linoleic, 14.2-33.3% oleic, 2.7-4.2% stearic and 12.0-16.6% palmitic acids. Murcia and Rincon [28] reported that pea oil contained 16.7-27.3% palmitic, 10.9-21.2% stearic, 14.9-21.7% oleic, 25.2-40.3% linoleic and 7.3-14.5% linolenic acids. In the discussion of the results obtained were probably due to temperature values during pea growth and the value of cotyledon/testa ratio during seed growth [28]. Also it is known that the enzymes involved in fatty acid biosynthesis depend on agroclimatic factors [29], and lipid content varies with climatic environmental conditions [30]. Solis et al. [31] reported that pea seed oil contained 3.67-9.01% palmitic, 3.44-7.31% stearic, 25.52-54.90% oleic, 21.38-44.78% linoleic and 6.01-14.01% linolenic acids. Kukavica et al. [32] reported that linoleic and oleic acids were the main unsaturated fatty acids of field pea, where as palmitic was main saturated fatty acid of field pea. Other researchers have found variable results for mineral contents and fatty acid

composition of pea seeds. There were also differences in the protein and oil contents of pea genotypes. Additionally, the seed specific parameters vary not only between kinds of seed but also for the same seed, due to different conditions in climate, soil, and harvesting [33,34]. The current study contributes to the available information concerning the composition of several pea genotypes grown in Turkey.

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REFERENCES

- [1] Iqtidar A., Akbar S., Khatoun S., Chemical Composition and Nutritional Evaluation of Peas Grown in NWFP (Pakistan), *J. Sci. Technol.*, **6**: 114-120 (1982).
- [2] Urbano G., Aranda P., Gomez-Villalva E., Nutritional Evaluation of Pea (*Pisum sativum* L.) Protein Diets After Mild Hydro Thermal Treatment and with and Without Added Phytase., *J. Agric.Food Chem.*, **51**: 2415-2420 (2003).

- [3] Jabeen T., Iqbal P., Khalil I.A., [Amino Acid and Mineral Composition of Pea Cultivars Grown in Peshawar](#), *Pak. J. Agric. Res.*, **9**: 2- (1988).
- [4] Ashraf M.I., Pervez M.A., Amjad M., Ahmad R., Ayub M., [Qualitative and Quantitative Response of Pea \(*Pisum sativum* L.\) Cultivars to Judicious Applications of Irrigation with Phosphorus and Potassium](#), *Pak. J. Life Soc. Sci.*, **9**(2): 159-164 (2011).
- [5] Woźniak A., Soroka M., Stepniowska A., Makarski B., [Chemical Composition of Pea \(*Pisum sativum* L.\) Seeds Depending on Tillage Systems](#), *J. Elem. Sci.*, 1143-1152 (2014).
- [6] Friedman M., [Nutritional value of Proteins from different food sources](#), A Review. *J. Agric. Food Chem.*, **44**: 6-29 (1996).
- [7] Bastianelli D., Grosjean F., Peyronnet C., Duparque M., Regnier J.M., [Feeding value of pea \(*Pisum sativum* L.\) 1. Chemical Composition of Different Categories of Pea](#), *Anim. Sci.*, **67**: 609-619 (1998).
- [8] Stanek M., Zduńczyk Z., Purwin C., Stefan Florek F., [Chemical Composition and Nutritive Value of Seeds of Selected Pea Varieties](#), *Vet. Ir Zootechnik*, **28**(50): 71-73 (2004).
- [9] Ceyhan E., Harmankaya M., Avcı M.A., Effects of Sowing Dates and Cultivars on Protein and Mineral Contents of Bean (*Phaseolus vulgaris* L.), *Asian J. Chem.* **20**(7): 5601-5613 (2008).
- [10] George R.A.T., Stephens R.J., Varis S., "The Effect of Mineral Nutrients on the Yield and Quality of Seeds in Tomato", In: Seed production (Ed. Hebblethwaite P.D), pp: 561-567 (1980).
- [11] Savage G.P., Deo S. The nutritional Value of Peas (*Pisum sativum*). A Literature Review, *Nutr. Abst. Rev. (Ser. A)*, **59**: 65-88 (1989).
- [12] Coxon D.T., Davies D.R., [The Effect of Therandrloci on the Lipid Content of the Seed of *Pisum sativum*](#), *Theor. Appl. Genet.*, **64**: 47-50 (1982).
- [13] Mccurdy S.M., Drake S.R., Swanson B.G., Leung H.K., Powers J.R., [Influence of Cultivars, Soak Solution, Blanch Method and Brine Composition on Canned Dry Pea Quality](#), *J. Food Sci.*, **48**: 394-399 (1983).
- [14] Akcin A. "Yemeklik Tane Baklagiller", Selcuk University Faculty of Agriculture Konya, Publication No. 8, 41-189 (1988).
- [15] AACC. International. "Method 46-30.01. Crude Protein - Combustion Method". In: *Approved Methods of Analysis 11th AACC International*: St. Paul, MN, USA (1999).
- [16] Matthaus B., Özcan M.M. [Quantification of Fatty Acids, Sterols and Tocopherols Turpentine \(*Pistacia terebinthus* Chia\) Wild Growing in Turkey](#), *J. Agric. Food Chem.*, **54**: 7667-7671 (2006).
- [17] Hişil Y., "Instrumental Analysis Techniques" (Eng. Fac. Publ. 55). Ege University, Bornova -İzmir. (in Turkish), (1998).
- [18] Skujins S., Hand book for ICP-AES (Varian-Vista). "A Short Guide To Vista Series ICP-AES Operation", Varian Int. AGşZug. Version 1.0. pp 29. Switzerland (1998).
- [19] Püskülcü H., İkiz F., "Introduction to Statistics", Bilgehan Presss, p 333, Bornova, Izmir, Turkey (1989) [in Turkish].
- [20] Harmankaya M., Özcan M.M., Karadaş S., Ceyhan E., [Protein and Mineral Contents of Pea \(*Pisum sativum* L.\) Genotypes Grown in Central Anatolian Region of Turkey](#), *South Western J. Hort. Biol. Environ.*, **1**(2): 159-165 (2010).
- [21] Wang N., Daun J.K., [Effect of Variety and Crude Protein Content on Nutrients and Certain Antinutrients in Field Peas \(*Pisum sativum*\)](#), *J. Sci. Food Agric.*, **84**: 1021-1029 (2004).
- [22] Welch R.W., Griffiths D.W., [Variation in the Oil Content and Fatty Acid Composition of Field Beans \(*Vicia faba*\) and Peas \(*Pisum* spp\)](#), *J. Sci. Food Agric.*, **35**: 1282-128 (1984)
- [23] Yoshida H., Tomiyama Y., Tanaka M., Mizushima Y., [Characteristic profiles of Lipid Classes, Fatty Acids and Triacylglycerol Molecular Species of Peas \(*Pisum sativum* L.\)](#), *Eur. J. Lipid Sci. Technol.*, **109**(6): 600-607 (2007).
- [24] Coxon D.T., Wright D.J., [Analysis of Pea Lipid Content by Gas Chromatographic and Microgravimetric Methods. Genotype Variation in Lipid Content and Fatty Acid Composition](#), *J. Sci. Food Agric.*, **36**: 847-856 (1985).
- [25] Ryan E., Galvin K., O'Connor T., Maguire A., O'Brien N., [Phytosterol, Squalene, Tocopherol Content and Fatty Acid Profile of Selected Seeds, Grains, and Legumes](#), *Plant Foods Hum. Nutr.*, **62**(3): 85-91 (2007).

- [26] Srivastava R.P, Kumar L., Dixit G.P., [Nutritional Composition and Fatty Acid Profile of Important Genotypes of Field Pea \(*Pisum sativum* ssp. *Arvense*\)](#), *J. Food Legumes*, **22**(2): 115-117 (2009).
- [27] Zhigacheva I., Burlakova E., Misharina T., Terenina M., Krikunova N., Generozova I., Shugaev A., Saidgarey Fattakhov S., [Fatty Acid Composition and Activity of the Mitochondrial Respiratory Chain Complex I of Pea Seedlings Underwater Deficit](#), *Biologija*, **59**: 241–249 (2013).
- [28] Murcia M.A., Rincon F., [Fatty Acid Composition of Pea \(*Pisum sativum* L. var. *Citrina*\) During Growth](#), *Grasas y Aceites*, **42**: 444-449 (1991).
- [29] Harwood J.L., Stump P.K., [Fat Metabolism in Higher Plants. XI. Synthesis of Fatty Acids in the Initial Stage of Seed Germination](#), *Plant Physiol.*, **46**: 500-508 (1970).
- [30] Worthington R.E., Hammos R.O., Allison J.R., [Varietal Differences and Seasonal Effects on Fatty Acid Composition and Stability of Oil from 82 Peanut Genotypes](#), *J. Agric. Food Chem.*, **20**: 727-732 (1972).
- [31] Solis M.I.V., Patel A., Orsat V., Singh J., Mark Lefsrud M. [Fatty Acid profiling of the Seed Oils of Some Varieties of Field Peas \(*Pisum sativum*\) by RP-LC/ESI-MS/MS: Towards the Development of an Oil Seed Pea](#), *Food Chem.*, **139**: 986-993 (2013).
- [32] Kukavica B., Quartacci M.F., Veljovic-Jovanovic S., Navari-Izzo F. [Lipid Composition of Pea \(*Pisum sativum* L.\) and Maize \(*Zeamays* L.\) Root Plasma Membrane and Membrane-Bound Peroxidase and Superoxide Dismutase](#), *Arch. Biol. Sci. Belgrade*, **59**(4): 295-302 (2007).
- [33] Stein W., Glaser F.W., [Continuous Solvent Extraction of Sunflower, Seed, Groundnuts, Palm Kernels, Rape Seed, and Copra](#), *J. Am. Oil Chem. Soc.*, **53**: 283-285 (1976).
- [34] Özcan M.M., Bağcı A., Dursun N., Gezgin S., Hamurcu M., Dumlupınar Z., Uslu N., [Macro and Micro Element Contents of Several Oat \(*Avena sativa* L.\) Genotype and Variety Grains](#), *Iran. J. Chem. Chem. Eng. (IJCCE)*, **36**: 73-79 (2017).