

## Engineered Algal Biochar for the Sequestration of $\text{Cu}^{2+}$ from Aqueous Solution

Thusabantu, Nathan; Munzeiwa Wisdom A.; Mudavanh Norman;

Mukaratirwa Muchanyereyi, Netai

Materials Research (MatRes) Group, Chemistry Department, Bindura University of Science Education,  
P Bag 1020, Bindura, ZIMBABWE

Chaukura, Nhamo\*<sup>+</sup>

Department of Physical and Earth Sciences, Sol Plaatje University, Kimberley, SOUTH AFRICA

**ABSTRACT:** The provision of safe drinking water in low-income countries is problematic due to high levels of pollution and the high cost of water treatment. While existing water treatment methods are efficient in removing most contaminants, they are expensive. ~~Adsorption~~ The adsorption methods may be a cheaper and more efficient alternative, given that feedstock for the fabrication of adsorbents, ~~are~~ is readily available, and they are easy to produce. The objective of this study was to synthesize and evaluate the performance of algae-derived adsorbents in removing  $\text{Cu}^{2+}$  from wastewater using batch experiments and fixed-bed columns. Algal biomass was pyrolyzed under limited oxygen to produce biochar (BC), which was separately activated using: (1) ferric chloride to form a  $\text{Fe}_2\text{O}_3$ -BC composite, and (2)  $\text{KMnO}_4$  and  $\text{H}_2\text{SO}_4$  through a modified Hummer's method to form HBC. Batch experimental data fitted well in both pseudo-first-order ( $r^2=0.965$ ) and pseudo-second-order ( $r^2=0.946$ ) kinetic models, and there was no significant difference ( $p=0.349$ ). The Yoon-Nelson ( $r^2=0.879$ ) and Thomas ( $r^2=0.891$ ) models adequately described the experimental data, while the Adams-Bohart model had a low fit ( $r^2=0.673$ ) in column studies. The results showed that the biosorbents were effective in removing  $\text{Cu}^{2+}$  from wastewater, with HBC having a higher affinity than  $\text{Fe}_2\text{O}_3$ -BC and BC. FTIR measurements after adsorption suggest that carbonyl groups played a key role in binding  $\text{Cu}^{2+}$  ions. Overall, valorizing algal biomass potentially helps ~~in solving to solve~~ the problem of algal blooms, while providing material for treating water. Further research should investigate the economic feasibility and up-scaling of the technology to field-scale.

**KEYWORDS:** Adsorption; Biochar; Breakthrough curve; Fixed-bed column modeling.

### INTRODUCTION

In recent years water pollution has become a cause for concern, especially in low-income countries where resources are limited and technology is lagging behind [1]. The increase

in pollution can be attributed to industrialization, increased mining and farming activities, and an increase in the population.

\* To whom correspondence should be addressed.

+E-mail: nhamo.chaukura@spu.ac.za & nchaukura@gmail.com  
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Pollutants such as industrial and agricultural chemicals,

and emerging contaminants such as pharmaceutical drugs, personal health care products, and surfactants pose a risk to human health as these may cause neurological disorders and reproductive problems [2]. In addition, artisanal mining operations have contributed to the accumulation of heavy metals such as Cd, Pb, Zn, Cr, and Cu in aquatic environments in low-income countries [3]. These metals are also introduced into the aquatic environment via industrial effluents, commercial mining processes, and plumbing in drinking water distribution systems [4]. Heavy metals are toxic and may cause nerve and tissue damage [5]. Of these, Cu is widely distributed, and exposure to excessive amounts can cause serious health problems because of its bioaccumulation and toxic effects [6]. It is therefore important to reduce exposure through efficient water treatment methods.

While existing water treatment methods such as coagulation, membrane filtration, and photocatalysis are effective, they are usually expensive, and difficult to implement. In contrast, traditional methods such as sand filtration are inefficient in removing chemical contaminants and pathogens [7, 8]. Conversely, adsorption methods using biochar have many advantages which include ease of design, low operations cost, and it is largely an environmentally friendly method [8]. Biochar is an ideal adsorbent since it has high porosity, large surface area, high thermal stability, an abundance of functional groups, and good adsorption capacity. A variety of materials can be used as adsorbent feedstock depending on the availability and cost of haulage. The most commonly used adsorbents are zeolites, activated carbon, and silica gel [9, 10]. Precursors used in the commercial production of biochar are non-renewables like coke, coal, and lignite, these are not sustainable and are relatively expensive materials [11]. Exploration for cheaper and naturally occurring feedstock such as biomass is, therefore, necessary to enhance efficiency and reduce water treatment costs.

A pyrogenic adsorbent derived from biomass, biochar, is a cost-effective and environmentally friendly adsorbent with a highly porous structure and is rich in oxygen-containing functional groups [9, 12-14]. The physicochemical properties of biochar such as surface charge, functional groups, morphology, and textural properties can be modified to improve its sorptive performance in water treatment [15]. Consequently, biochar has great promise in

water treatment. Feedstock for biochar fabrication includes agro-waste, municipal sludge, and forestry residues [1, 16]. With a high biomass yield, algae is a lucrative feedstock in biochar production, as it is renewable, requires no specialized inputs for cultivation, and does not compete with food sources [14]. Moreover, the use of algae as biochar feedstock could potentially eliminate the problems associated with algal blooms [14]. Algal biochar can be modified to enhance its adsorption capabilities. According to previous studies, biochar derived from algae has a high heavy metal removal capacity compared to other biochars from woody material [17]. In addition, algal biochar has been used in the removal of anionic and cationic pollutants [14]. Chemical activation of algal biochar using the Hummers method can be used to increase the amount of oxygen-containing functional groups [18]. This method involves chemical oxidation using  $\text{KMnO}_4$  and  $\text{H}_2\text{SO}_4$ , which increases the surface area by introducing C=O, C-O, and COOH groups, resulting in an improved adsorption performance [19, 20].

Although several studies have reported the removal of  $\text{Cu}^{2+}$  using biochar derived from agricultural wastes and plant residues [21-23], the efficiency of most biochars is limited by surface precipitation between  $\text{CO}_3^{2-}$  or  $\text{PO}_4^{3-}$  and  $\text{Cu}^{2+}$  [24]. While previous studies have used biochar sorbents for removing heavy metals in aquatic systems, a limited number of studies have reported the use of biochar derived from algae for the removal of  $\text{Cu}^{2+}$  from wastewater. This study seeks to explore the sorptive performance of algal biochar and how it can be modified to improve efficiency in removing  $\text{Cu}^{2+}$  from synthetic wastewater. The specific objectives were: (1) to synthesize and characterize chemically modified biosorbents from algae, and (2) to use batch experiments and fixed-bed columns in evaluating the sorptive performance in removing  $\text{Cu}^{2+}$  from synthetic wastewater. The modification techniques used are time- and cost-efficient, and remove a range of divalent cations from aquatic systems.

## EXPERIMENTAL SECTION

### Preparation of adsorbents from algae

#### Biochar

Algae were collected from a river and washed thoroughly to remove fine soil particles and dirt particles

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before being sun-dried for 24 h. The material was further dried in an oven for 2 h at 60 °C. After cooling to room temperature, the algae was ground using a mortar and pestle **and** then passed through a 0.25-2 mm sieve. Ground algae were placed in porcelain crucibles, covered with aluminum foil, and placed in airtight canisters to minimize contact with air. These were **pyrolysed-pyrolyzed** at 400 °C furnace at a heating rate of 7 °C/min for 2 h. The biochar was cooled slowly to room temperature, labeled BC, and stored in a self-sealing polythene bag

#### *Fe<sub>2</sub>O<sub>3</sub>-BC*

The BC was immersed in a 40 g/100 mL solution of FeCl<sub>3</sub>.6H<sub>2</sub>O in the ratio 1:3 (m/v) for 3 h. The mixture was then filtered, and the residue dried in an oven for 2 h at 80 °C. The dry material was pyrolyzed at 400 °C for 3 h. The composite material formed was allowed to cool gradually, labeled Fe<sub>2</sub>O<sub>3</sub>-BC, and stored in a self-sealing polythene bag for further use.

#### *Hummers biochar*

To synthesize Hummers biochar (HBC), a 15 g sample of BC was mixed with 30 mL of 1 M KMnO<sub>4</sub>. To this, concentrated H<sub>2</sub>SO<sub>4</sub> (10 mL) was added dropwise. The mixture was agitated for 10 min and allowed to stand for 1 h. The slurry formed was filtered and washed several times with deionized water to remove residual acid. The resulting material was pyrolyzed at 400 °C for 2 h, after which it was cooled to room temperature and stored in a self-sealing polythene bag for further use.

#### *Characterization of biosorbents*

##### *Determination of pH, pHzpc, ash content, and cation exchange capacity*

The measurement of pH was performed on 1 g of adsorbent suspended in deionized water (10 mL) and agitated for 10 min. The pH drift method was used to measure pHzpc [25]. The ash content of the adsorbents was determined by igniting 1g of adsorbent in a muffle furnace at 600 °C for 2 h and measuring the weight of the resulting ash. To determine the cation exchange capacity (CEC), 1 g of adsorbent sample was mixed with 50 mL of 1 M HCl in a 100-mL beaker, and the mixture was placed on a shaker for 2 h. The mixture was filtered and washed several times with deionized water. The sample was then added to 50 mL of 1 M ammonium

acetate and shaken for 1 h, and filtered. The filtrate was titrated with 0.5 M NaOH solution until the endpoint, and CEC calculate (Eq. (1)):

$$CEC = \frac{C_{NH_4^+}}{M} V \quad (1)$$

Where *C* is the concentration of NH<sub>4</sub><sup>+</sup>, *V* is the volume of the filtrate, and *M* is the mass of the sample.

#### *Surface functional groups, surface morphology, and crystallinity*

The surface functional groups on the adsorbents were determined using a Fourier Transform Infra-Red (FT-IR) spectrometer (Nicolet, iS5) operated at a resolution of 2 cm<sup>-1</sup>, an average of 16 scans, in the range 400-4000 cm<sup>-1</sup>. Surface morphologies were determined on gold-coated samples using a Scanning Electron Microscope (SEM) (JEOL, JSM-IT300, Japan) at 50 μm magnification, and crystalline properties were determined in the 2θ range 4-55° using an X-ray diffractometer (Bruker, D2 Phaser) equipped with a Cu-Kα radiation source (λ=1.54 Å).

#### *Batch studies*

##### *Adsorption equilibrium*

Batch adsorption experiments were performed in 250 mL Erlenmeyer flasks with a 50 mg/L stock solution Cu<sup>2+</sup>. Working solutions (50 mg/L, 25 mg/L, and 12.5 mg/L) were prepared from the stock solution by serial dilution. Each of the biosorbent materials (1 g) was mixed with 50 mL Cu<sup>2+</sup> solution, and the mixtures were agitated for 90 min while monitoring the concentration of residual Cu<sup>2+</sup> at 5 min intervals using a UV-Vis spectrophotometer (Thermo Scientific, Genesys 10S) at 836 nm wavelength. All experiments were performed at room temperature and a neutral pH of 7.0 ± 0.1, controlled by 0.1M solutions of NaOH and HCl. The amount of contaminant adsorbed at equilibrium (*q<sub>e</sub>*) was calculated (Eq. 2):

$$q_e = \frac{C_o - C_e}{m} V \quad (2)$$

Where *C<sub>o</sub>* is the initial concentration of contaminant, *C<sub>e</sub>* equilibrium concentration of contaminant, *m* is the mass of adsorbent and *V* is the volume of solution.

##### *Kinetics*

The kinetics of the adsorption process were followed

by monitoring the change in concentration at 15 min intervals over a period of 75 min at a temperature of 25 °C. Pseudo-First-Order (PFO) (Eq. (3)) and Pseudo-Second-Order (PSO) (Eq. (4)) equations were used to describe the kinetics [5].

$$\ln(q_e - q_t) = \ln q_e - k_1 t \quad (3)$$

$$\frac{t}{q_t} = \frac{t}{q_e} + \frac{1}{k_2 q_e^2} \quad (4)$$

Where  $q_e$  and  $q_t$  are the amount of  $\text{Cu}^{2+}$  adsorbed at equilibrium and at time  $t$ , respectively,  $k_1$  and  $k_2$  are the PFO and PSO constants, respectively.

#### Fixed-bed column experiments

Glass columns with a diameter of 1 cm were each packed with a cotton wool plug, followed by a layer of adsorbent. Each layer of adsorbent consisted of 7 g of adsorbent for the 7-cm column, 2.5 g for the 3-cm column, and a 2 cm layer of acid-washed sand at the top. The concentration of  $\text{Cu}^{2+}$  in the effluent was measured using spectrophotometry every 50 min. Breakthrough curves, which relate the ratio of inlet concentration to the outlet concentration ( $C/C_0$ ) to time [26], were used to describe the removal efficiency of the columns (Eq. (5)):

$$q_t = \frac{Q C_i}{1000} \int_0^t \left(1 - \frac{C_t}{C_i}\right) dt \quad (5)$$

Where  $q_t$  is the amount of adsorbate after time  $t$  (min),  $Q$  is the flow rate (mL/min),  $C_i$  and  $C_t$  are the inlet and outlet concentrations, respectively (mg/L).

Experimental data were fitted onto mathematical models and used to describe the efficiency indices of the column, e.g., removal capacity and exhaustion time. The Thomas model (Eq. 6) assumes the rate of adsorption is controlled by surface interactions between the adsorbate and unoccupied active sites of the adsorbent [27]. It also assumes Langmuir kinetics without axial dispersion [27, 28].

$$\ln\left(\frac{C_0}{C_e} - 1\right) = \frac{K_{Th} q_0 m}{Q} - \frac{K_{Th} C_0 V}{Q} \quad (6)$$

Where  $C_0$  is the initial concentration,  $C_e$  is the equilibrium concentration,  $q_0$  is the maximum adsorption capacity (mg/g),  $m$  is the mass of the adsorbent (g),  $V$  is the throughput volume (mL), and  $K_{Th}$  is the Thomas constant (mL/min/mg).

The Yoon-Nelson model (Eq. (7)) assumes that the rate of decrease in adsorption probability of each adsorbate is proportional to the probability of adsorption and adsorbate breakthrough on the adsorbent [29]:

$$\ln\left(\frac{C_t}{C_0 - C_t}\right) = K_{YN} \tau - \tau K_{YN} \quad (7)$$

Where  $C_t$  is the concentration of  $\text{Cu}^{2+}$  at time  $t$ ,  $K_{YN}$  is the rate constant ( $\text{min}^{-1}$ ), and  $\tau$  (min) is the time required for 50% adsorbate breakthrough.

The parameters  $K_{YN}$  and  $\tau$  are obtained by plotting  $\ln[C/(C_0 - C)]$  against  $t$ . In the Adams-Bohart model (Eq. (8)), the rate of adsorption is proportional to the residual capacity of the adsorbent and the concentration of the adsorbate [27]. The model was used to describe the first part of the breakthrough curve.

$$\ln\left(\frac{C_t}{C_0}\right) = K_{AB} C_0 t - K_{AB} N_0 \frac{h}{U_0} \quad (8)$$

Where  $K_{AB}$  is the rate constant,  $U_0$  is the linear velocity,  $h$  is the bed height, and  $N_0$  is the saturation concentration.

## RESULTS AND DISCUSSION

### Characteristics of adsorbents

#### Physico-chemical characteristics

The pH data show that HBC is the most acidic, followed by  $\text{Fe}_2\text{O}_3\text{-BC}$ , then BC which is alkaline (Table 1). In agreement with previous reports, the activation processes used in the synthesis of HBC and  $\text{Fe}_2\text{O}_3\text{-BC}$  introduced more acidic functional groups such as C-O, and COOH [30]. Moreover, thermal treatment might have led to the degradation of alkaline functional groups [31]. The pH affects the electrostatic interaction between adsorbate and adsorbent by influencing the surface charge [17]. Previous studies reported that, as the initial solution pH increases from 3 to 5.5, the sorption of  $\text{Cu}^{2+}$  on biochar increases, and decreases thereafter due to the formation of the hydroxide [6]. The surface charge of the adsorbent becomes less positive with increasing pH, and thus the affinity for cations increases [19, 32].

The  $\text{pH}_{zpc}$  value was greatest for  $\text{Fe}_2\text{O}_3\text{-BC}$  and least for BC. This can be attributed to the different surface functional groups introduced by the activation methods used in the synthesis of biosorbents. While HBC was activated with  $\text{H}_2\text{SO}_4$ ,  $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$  was used in producing  $\text{Fe}_2\text{O}_3\text{-BC}$ . The basic nature of  $\text{Fe}_2\text{O}_3$  resulted in the higher  $\text{pH}_{zpc}$

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of the adsorbent. The number of acidic functional groups was lower than the basic groups as indicated

**Table 1: Physicochemical properties of biosorbents.**

Adsorbent	pH	pH <sub>zpc</sub>	CEC (meq/100 g)	Ash%
BC	8.22±0.1	8.38± 0.1	36± 1.5	39.9±2.4
HBC	4.28± 0.1	8.10± 0.1	48± 2.0	34.7±0.1
Fe <sub>2</sub> O <sub>3</sub> -BC	5.84± 0.2	8.41± 0.1	41± 1.5	38.9±0.4

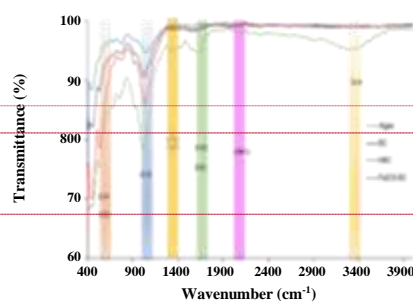
by the generally high pH<sub>zpc</sub> values. The pH<sub>zpc</sub> is the pH at which the number of positive and negative charges are equal. Below this pH there are more positive functional groups, and above this pH there are more negative functional groups [33]. The CEC of the adsorbents ranged from 36-48 meq/100 g, which was comparable to those in the literature (29-41 meq/100 g) [34]. Having the highest CEC, HBC was expected to adsorb more cationic solutes, while BC would be the least effective. CEC is a measure of the number of cations that can be adsorbed on an adsorbent [10].

#### Surface functional groups

The FT-IR spectra of biosorbents had bands appearing on similar positions and wavelengths as reported in previous studies [11, 35]. Materials BC, HBC, and Fe<sub>2</sub>O<sub>3</sub>-BC showed common peaks corresponding to -C-O stretch vibrations at approximately 1030 cm<sup>-1</sup> (shaded blue) (Fig. 1), O-H band at 3400 cm<sup>-1</sup> (orange), acyl amino acids, C=C and C=O stretch at 1550-1620 cm<sup>-1</sup> (green) and C-H, C-C bending vibrations at 740-780 cm<sup>-1</sup> (red). Algal biomass had a pronounced peak at 1400 cm<sup>-1</sup> (yellow), showing the presence of aromatic rings. The pyrolysis process modified the surface functional groups as evidenced by FT-IR spectra of BC. Notably, the O-H band (3300 cm<sup>-1</sup>) disappeared, while the C-H and C-Cl peaks at around 740 cm<sup>-1</sup> diminished. The activation of BC to HBC introduced -C≡C- stretch vibrations at 2175 cm<sup>-1</sup> (purple), H-C=O vibrations (2638 cm<sup>-1</sup>), and O-H vibrations from alcohols, carboxylic acids, and phenols (2329 cm<sup>-1</sup>) [36]. The Fe-O bond in Fe<sub>2</sub>O<sub>3</sub>-BC was evident at 532 cm<sup>-1</sup>, and the peak at 447 cm<sup>-1</sup> in HBC was assigned to the Mn-O bond, confirming the presence of Mn on the engineered biochar. Overall, the presence of both polar and non-polar functional groups on the surface of the adsorbents suggested they can interact with Cu<sup>2+</sup> through chemisorption or physisorption [14].

#### Surface morphology and crystallinity

The SEM images show that algal biomass has spongy



**Fig. 1: FT-IR spectra of Algae, BC, HBC, and Fe<sub>2</sub>O<sub>3</sub>-BC.**

heterogeneous surface (Fig. 2a), which changed after pyrolysis to a rough granular surface with more pronounced porosity evident on the surface of BC (Fig. 2b). While HBC consists of irregular cubic blocks with considerable inter-particle space (Fig. 2c), Fe<sub>2</sub>O<sub>3</sub>-BC had a honeycomb structure with regular pores (Fig. 2d). The surface heterogeneity and availability of pores suggest the adsorbents were suitable for adsorption [37]. From the SEM images, the surface of Fe<sub>2</sub>O<sub>3</sub>-BC had more dilated pores, therefore the material can potentially accommodate more Cu<sup>2+</sup> ions. The granular surface of BC creates more active sites compared to the large boulders of HBC.

The XRD spectra show that BC is largely amorphous, while HBC and Fe<sub>2</sub>O<sub>3</sub>-BC have distinct crystalline phases (Fig. 3). The broadening and low intensity of the BC spectrum are indicative of the low graphitic nature of BC, characteristic of amorphous carbon [38]. The peak at 2θ=20.3° for HBC and Fe<sub>2</sub>O<sub>3</sub>-BC, and a broad peak at 20° for BC corresponds to the 001 planes of amorphous carbon [39]. The HBC showed crystalline phase peaks at 2θ = 28.1°, 28.9°, 34.9°, 36°, 37°, and 44° due to MnO<sub>x</sub> crystal planes [40,

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41]. However, some of the  $MnO_x$  peaks were not very intense suggesting limited bulk loading of the  $KMnO_4$ . Previous studies suggest that the presence of  $MnO_x$

particles on  $KMnO_4$ - $KMnO_4$ -treated biochar could contribute

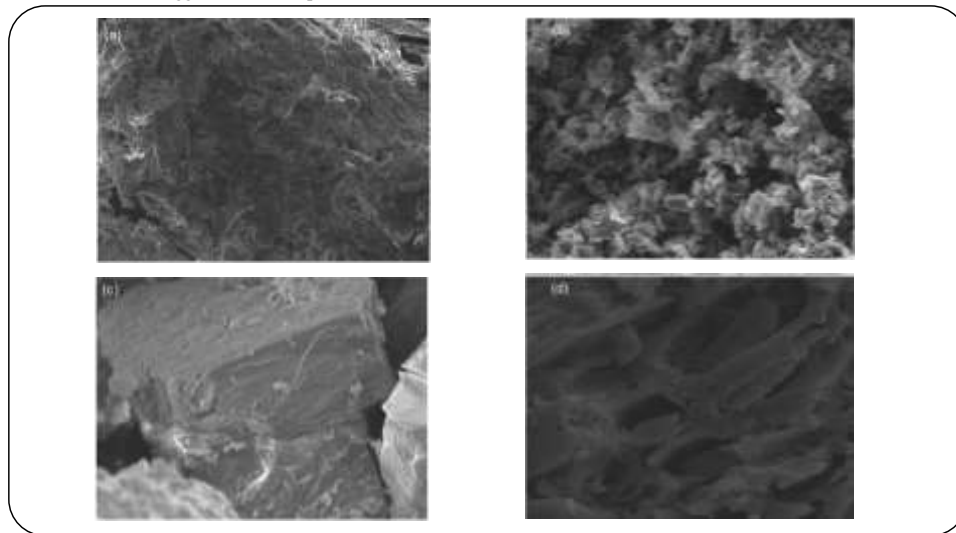


Fig. 2: SEM images of (a) algae (b) BC, (c) HBC, and (d)  $Fe_2O_3$ -BC at 50  $\mu m$  magnification.

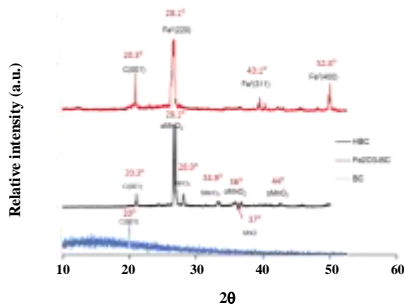


Fig. 3: XRD spectra for BC, HBC, and  $Fe_2O_3$ -BC.

to adsorption of pollutants and cause the generation of hydroxyl groups [42, 43]. For  $Fe_2O_3$ -BC, typical  $Fe_2O_3$  diffraction peaks were observed at  $2\theta$  values 28.1°, 43.1°, 52.8° assigned to 220, 311, 400 crystal planes of  $Fe_2O_3$ , respectively [39].

#### Kinetic studies

BC had the highest adsorbate uptake ( $q_e$ ) (3.12 mg/g),

followed by HBC (2.49 mg/g), then  $Fe_2O_3$ -BC (2.14 mg/g) (Fig. 4a). This could be because BC had a slightly more negative surface charge ( $pH_{zpc}=8.38$ ) which attracted  $Cu^{2+}$ , and the granular surface of BC availed more active sites for adsorption. From the higher number of oxygenated groups which potentially bind  $Cu^{2+}$  ions, HBC was expected to have the highest  $q_e$ . However, large granules could have reduced the surface area and the number of binding sites. The rate constant,  $k$ , showed the adsorption process on BC ( $5.31 \times 10^{-3} g/(mg \text{ min})$ ) was slower than on HBC ( $1.22 \times 10^{-2} g/(mg \text{ min})$ ) (Fig. 4b). Surface carboxylic and hydroxyl groups may be blocked by  $Fe_2O_3$  particles, making them inaccessible for  $Cu^{2+}$  adsorption [10].

From the experimental data, the  $q_e$  values were significantly ( $p=0.05$ ) greater than for the PSO (2.47 mg/g) for the PFO model (1.82 mg/g) (Table 2). There was no significant difference between the  $r^2$  values of PFO and PSO plots ( $p<0.05$ ), suggesting adsorbate-adsorbent interactions involved both physisorption and chemisorption [44]. Chemical interactions between the  $Cu^{2+}$  and polar functional groups (C=O, O-H, N-H, H-C=O, Fe-O) on the surface of the adsorbent are expected.

### Fixed-bed column experiments

#### Breakthrough curves

In 3-cm columns, the breakthrough times were in the order: BC<Fe<sub>2</sub>O<sub>3</sub>-BC<HBC (Fig. 5a), and increased in the order: Fe<sub>2</sub>O<sub>3</sub>-BC<BC<HBC for 7-cm columns (Fig. 5b).

Table 2: Kinetic parameters of adsorption of Cu<sup>2+</sup> on BC, HBC, and Fe<sub>2</sub>O<sub>3</sub>-BC.

Adsorbent	Kinetic model	Parameter	Value
BC	PFO	q <sub>e</sub> (mg/g)	1.942
		k <sub>1</sub> (min <sup>-1</sup> )	4.55×10 <sup>-2</sup>
		r <sup>2</sup>	0.859
	PSO	q <sub>e</sub> (mg/g)	3.117
		k <sub>2</sub> (g/(mg min))	5.31×10 <sup>-3</sup>
		r <sup>2</sup>	0.875
HBC	PFO	q <sub>e</sub> (mg/g)	1.719
		k <sub>1</sub> (min <sup>-1</sup> )	0.542
		r <sup>2</sup>	0.956
	PSO	q <sub>e</sub> (mg/g)	2.49
		k <sub>2</sub> (g/(mg min))	1.22×10 <sup>-2</sup>
		r <sup>2</sup>	0.812
Fe <sub>2</sub> O <sub>3</sub> -BC	PFO	q <sub>e</sub> (mg/g)	1.829
		k <sub>1</sub> (min <sup>-1</sup> )	4.3×10 <sup>-2</sup>
		r <sup>2</sup>	0.993
	PSO	q <sub>e</sub> (mg/g)	2.14
		k <sub>2</sub> (g/(mg min))	1.36×10 <sup>-2</sup>
		r <sup>2</sup>	0.941

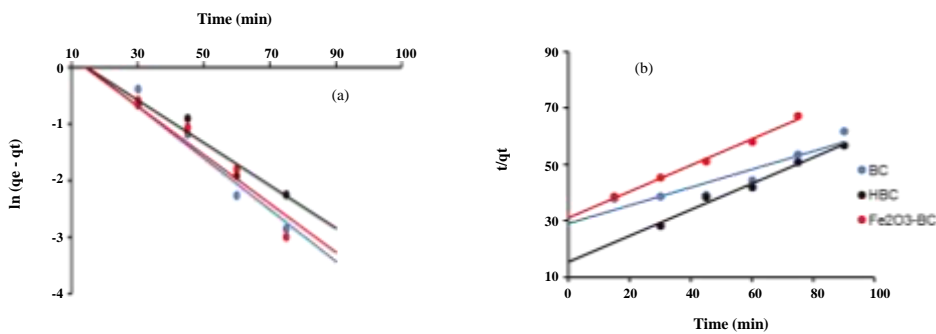


Fig 4: (a) Pseudo-first-order pseudo plot, and (b) pseudo-second-order plot for Cu<sup>2+</sup> adsorption.

The results show that the breakthrough time varies directly with the bed height of the column. The performance

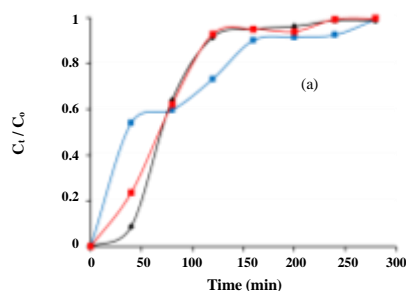
of a column is evaluated using breakthrough curves i.e. the time to reach a breakthrough and the shape of the curve [26].

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Further analysis of the curves was performed using the Thomas (Table S1), Yoon-Nelson (Table S2), and Adams-Bohart (Table S3) models.



#### Mathematical modeling of column studies

The rate constant for the Thomas model ( $K_{Th}$ ) was

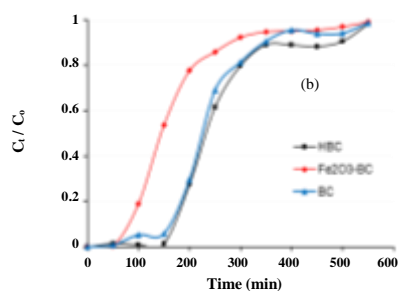


Fig. 5: Breakthrough curves for the adsorption of  $\text{Cu}^{2+}$  using (a) a 3-cm bed height, and (b) a 7-cm bed height.

dependent on the amount of adsorbent/height of the column and the flow rate (Fig. 6a). Columns of 3 cm bed height had significantly ( $p=0.05$ ) higher  $K_{Th}$  values ( $3.30\text{--}5.17 \times 10^{-4}$  mL/(min.mg)) compared to the 7-cm bed height ( $2.07\text{--}3.34 \times 10^{-4}$  mL/(min.mg)) (Table S1). The highest  $K_{Th}$  value ( $5.17 \times 10^{-4}$  mL/(min.mg)) was observed in  $\text{Fe}_2\text{O}_3\text{-BC}$  at 3 cm bed height, while the least value ( $2.07 \times 10^{-4}$  mL/(min.mg)) was observed for  $\text{Fe}_2\text{O}_3\text{-BC}$  bed height of 7 cm. The decrease of  $K_{Th}$  with increasing bed height was due to a reduced reaction rate caused by longer contact time for higher bed depth [45]. The equilibrium uptake ( $q_e$ ) was dependent on the bed height, with larger columns having significantly ( $p=0.05$ ) greater adsorption capacity (161.73 mg/g) compared to the shorter bed heights (142.03 mg/g). An increase in the bed height increased the active sites for adsorbate-adsorbent interactions. HBC had the highest  $q_e$  value (171.76 mg/g), probably due to negatively charged surface functional groups which have a higher affinity for the positively charged  $\text{Cu}^{2+}$  ions. For the Yoon-Nelson model,  $\tau$  was directly proportional to bed height and  $K_{YN}$ , and inversely proportional to the flow rate (Fig. 6c). When the flow rate increased the contact time reduced, resulting in less time for the adsorbate to traverse the pores and interact with the adsorbent [46]. HBC had the highest  $\tau$  value (299 min) relative to the other adsorbents (BC: 266 min,  $\text{Fe}_2\text{O}_3\text{-BC}$ : 205 min). The Adams-Bohart rate constant ( $K_{AB}$ ) was highest for HBC in 7-cm (0.028 L/mgmin) and BC in 3-cm (0.056 L/mgmin) columns (Fig. 6e,f). The lowest values

were observed in  $\text{Fe}_2\text{O}_3\text{-BC}$  for 7-cm column (0.026 L/(mg.min)) and HBC for 3-cm column (0.049 L/(mg.min)).

The value of  $K_{AB}$  increased with bed height and decreased with an increasing flow rate. The maximum values of the saturation concentration ( $N_0$ ) were observed in BC for 3-cm column (564.86 mg/L), and HBC for 7-cm column (276.56 mg/L). The  $N_0$  values varied linearly with the flow rate, and inversely with the bed height. The Adams-Bohart model assumes the rate of adsorption is dependent on the adsorption capacity that remains on the sorbent [47]. This model is used to describe the initial part of the breakthrough curve. In summary, the  $r^2$  values from the Thomas (0.891) and Yoon-Nelson (0.878) models were significantly higher than the values in the Adams-Bohart model (0.673). The values signify a good fitting of the mathematical models to the experimental data [45]. It, therefore, follows that adsorption is better described by the Thomas, and Yoon-Nelson models.

#### Mechanism of removal

For BC, a new peak was observed at  $540 \text{ cm}^{-1}$ , and was ascribed to the formation of a Cu-O bond (Fig. 7a). A similar peak was observed after the adsorption of  $\text{Cu}^{2+}$  on HBC (Fig. 7b). This showed the existence of chemical interactions, resulting in higher  $q_e$  values. A higher  $q_e$  value for HBC was also attributed to the presence of MnOx particles, which can form complexes with  $\text{Cu}^{2+}$  thus enhancing its sorption [41]. Notable was an increase

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in intensity and a red shift in the carbonyl group at  $1030\text{ cm}^{-1}$  (Fig. 7c) and an increase in intensity and a blue shift at  $1050\text{ cm}^{-1}$  (Fig. 7d). This suggested the mechanism involved chemical interactions between adsorbent and

adsorbate, thus following PSO kinetics. Carbonyl groups, being oxygen-carrying moieties, provide nascent lone electrons that can bind with toxic elements such as  $\text{Cu}^{2+}$  to form complexes [40].

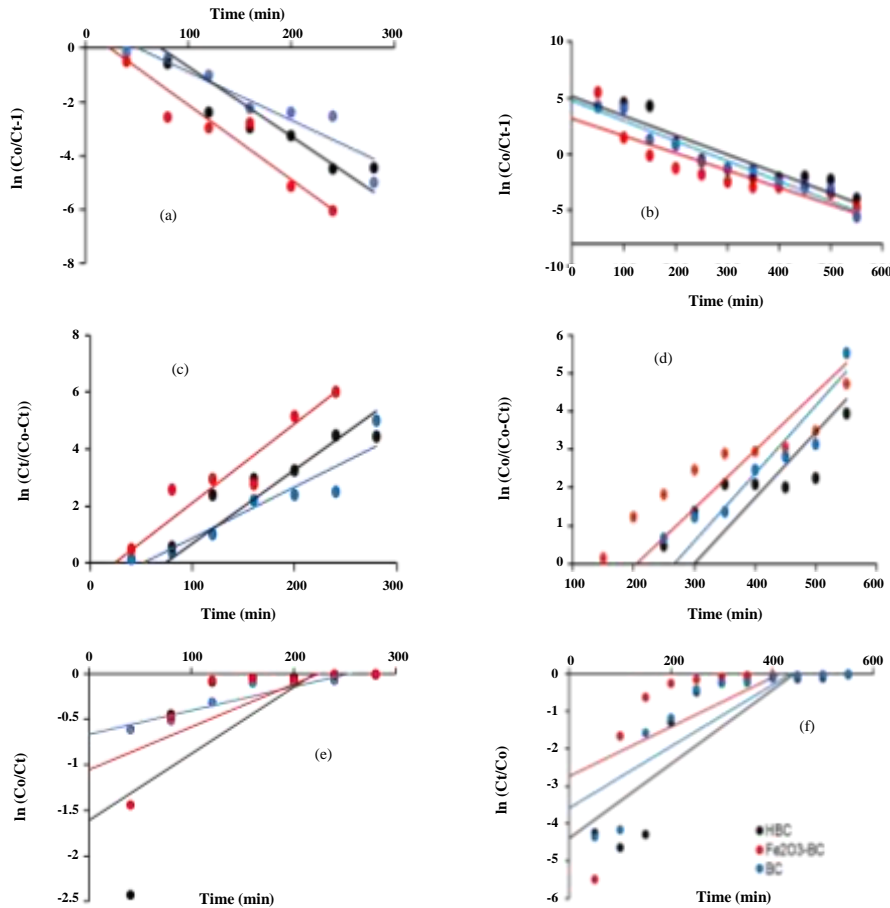


Fig. 6: (a) Thomas model for 3-cm column, and (b) 7-cm column, Yoon-Nelson model for (c) 3-cm column, and (d) 7-cm column; and Adams-Bohart model for (e) 3-cm column, and (f) 7-cm column

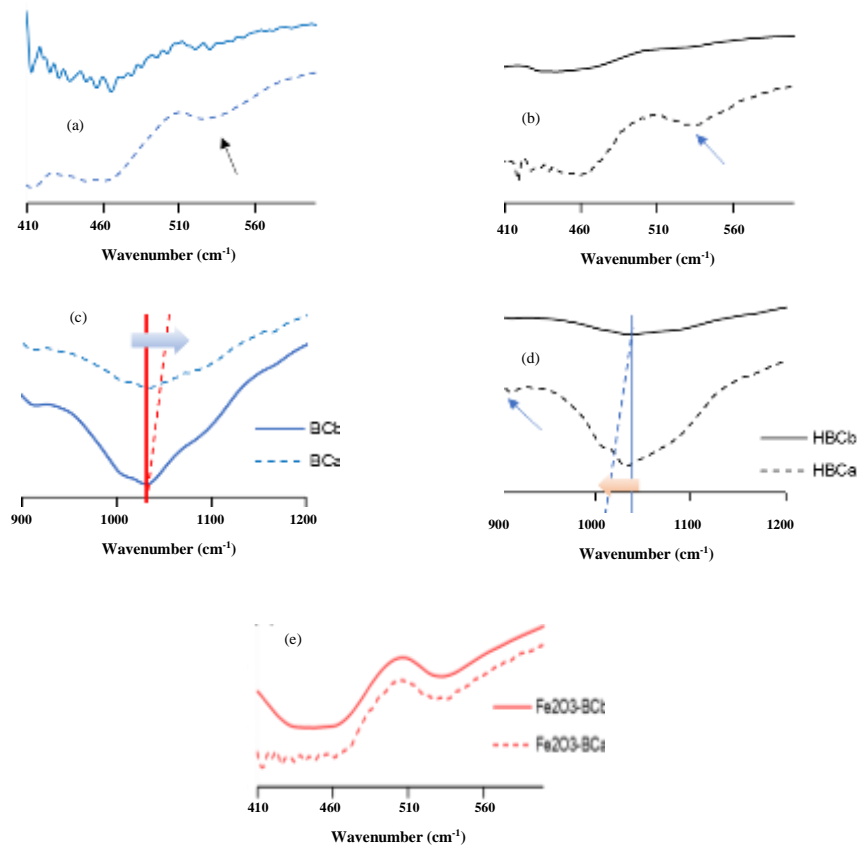
The FT-IR spectra before and after  $\text{Cu}^{2+}$  adsorption on  $\text{Fe}_2\text{O}_3\text{-BC}$  were similar (Fig. 7e). Chemical interactions between adsorbate and adsorbent were probably limited, hence  $\text{Fe}_2\text{O}_3\text{-BC}$  did not retain many  $\text{Cu}^{2+}$  ions, resulting in low  $q_e$  (2.14 mg/g). A comparison of the FT-IR spectra

for the adsorbents before and after adsorption gives an insight into the probable adsorption mechanism [36]. When an adsorbate interacts with a surface functional group on the adsorbent, the vibrational frequency is altered, leading to a shift in wavenumber, and sometimes the intensity

of a peak [14, 36]. Overall, the dominant mechanism for the adsorption process for all adsorbents may have involved chemical interactions between the adsorbent and adsorbate.

## CONCLUSIONS

The performance of algal-based biochar adsorbents in removing  $\text{Cu}^{2+}$  was evaluated using batch and column experiments. The key findings were:



**Fig 7: FT-IR spectra for BC, HBC, and  $\text{Fe}_2\text{O}_3\text{-BC}$  before and after adsorption of  $\text{Cu}^{2+}$ . The solid and dashed lines denote before and after adsorption, respectively.**

1- Batch experiments showed no significant distinction between the PFO and PSO kinetic models. The study showed that the adsorbents were effective as shown by the adsorption capacity values. BC was the most effective followed by HBC, then  $\text{Fe}_2\text{O}_3\text{-BC}$  in the removal of  $\text{Cu}^{2+}$  from the aqueous solution.

2- In column experiments, HBC was more effective than  $\text{Fe}_2\text{O}_3\text{-BC}$  and BC in the removal of  $\text{Cu}^{2+}$ . It was also

deduced that column height and flow rate affect the efficiency of adsorption.

FT-IR measurements after adsorption suggest that the carbonyl groups played a key role in the  $\text{Cu}^{2+}$  binding process. The adsorbents removed  $\text{Cu}^{2+}$  from an aqueous solution with higher concentrations than natural water bodies. The adsorption efficiencies of adsorbent materials were effective and exhibited great potential in water

treatment to acceptable standards. Vast amounts of algae and relative ease of design show the potential of the adsorbents to be adopted for large-scale studies. However, the sorbent materials can only be co-used with other

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