

# Spectral and Electrochemical Sensing Studies of Novel Tetradentate Schiff Bases, Having Enhanced Antifungal Activity, Containing Ferrocene and Aromatic Moieties Attached Imines

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**ABSTRACT:** Multi metal ion sensing unsymmetrical Schiff bases containing ferrocene group attached imine at one side and aromatic moiety connected azomethine on the other side have been synthesized. Titration studies coupled with a UV-Visible spectrophotometer expose the binding aptitude of new receptors. Development of MLCT band near 457 nm for the coordination of  $\text{Cu}^{2+}$  ions with new ligands is also noticed. Electrochemical studies of receptor solutions with added metal ions expose quasi-reversible processes by giving superfluous  $\Delta E_p$  values (146-161 mV, then the expected 59 mV). The concentration of metal ions required for effective sensing is calculated from the percentage amount of  $\Delta I_{pa}$  extracted from the  $I_{pa}$  data. Results obtained in computational molecular docking studies and in-vitro analysis invite more focused research by the pharmacist to develop new formulations for antifungal medicines.

**KEYWORDS:** Antifungal Compound; Binding Attitude; Cation Sensors; In-vitro; Molecular Docking.

## INTRODUCTION

Scientists in the field of Material science, Chemistry, Physics, and Biology looking for small abiotic molecules capable of binding discriminatively (reversibly or irreversibly) to different metal ions with a simultaneous noticeable change in the potential value of oxidation state or modification in the absorption and emission spectra of the employed component [1]. Chemosensors are tiny molecules that find application in different fields including clinical areas [2-5] for the reason that diminutive toxins and heavy metal ions cause health and environmental deterioration. Effluent from various industries, volcanic

explosion and farming activities introduce cations and anions which affect the environment as well as the health of humans [6] Low sensitivity and longer time duration are associated with instrumentation techniques utilized to identify hazardous metal ions [7]. Chemosensors primed from Schiff base reactions in the form of ligands and metal complexes have a wide range of applications in many fields including systematic analysis, biochemical investigations, and antimicrobial studies [8-10].

Mercury ions, the greatest toxic heavy and transition metal ions, found in the aquatic system can root impairment

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to the kidney, central nervous system, offspring of a mammal, and toddlers [11]. Though copper ions are involved in many metabolic activities of humans [12], unfettered intake of copper ions may lead to degeneration of the nervous system [13] and detriment to the liver & kidney [14]. Alternation in the function of proteins present in enzymes having metal ions like zinc and ferrous resulted from the ingestion of nickel ions [15]. Manganese ions toxicity comprises vacillation in brain function, low appetite, crippling of neural function, and vehement activity [16]. Protein metallothionein transmits  $Cd^{2+}$  ions to the kidney. The extended biotic half-life period of  $Cd^{2+}$  ions gives amassed toxic effects and affects the kidney and develops prostate cancer [17]. Accumulation of  $Pb^{2+}$  ions in blood diminishes hearing ability in children and postponements adolescents' sexual maturity [18].

An enormous amount of ligands and their transition metal ion coupled complexes have been synthesized and used as models to mimic the biological activities of enzymes [19]. Not only that, Schiff bases are used in diverged fields including analytical, pharmaceutical, medical, and environmental research field [20].

Herein, the synthesis of new synthetic unsymmetrical Schiff bases *N*'1-((*E*)-2-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide and *N*'1-((*E*)-2-hydroxy-5-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide possessing ferrocene component at one end and aromatic group at another end of the foremost structure is reported. The presence of azomethine (-CH=N) groups with lone pair of electrons on  $sp^2$  hybridized nitrogen atom is responsible for antifungal, antibacterial, and anticancer activities of Schiff base ligands and complexes [21]. Revelation of enhanced antifungal bustle rather than antibacterial activity of the newly prepared receptors alarms more pharmaceutical research to be carried out to bridge the gap between available antibacterial and antifungal commixture [22].

## EXPERIMENTAL SECTION

### Materials

Succinic anhydride, hydrazinehydrate, ferrocenecarboxaldehyde, 2-hydroxy-5-nitrobenzaldehyde, 2-nitrobenzaldehyde, and silica gel (Analytical grade) were purchased from E. Merck industry and used without further purification. Analytical grade  $HgCl_2$ ,  $Pb(OAc)_2$ ,

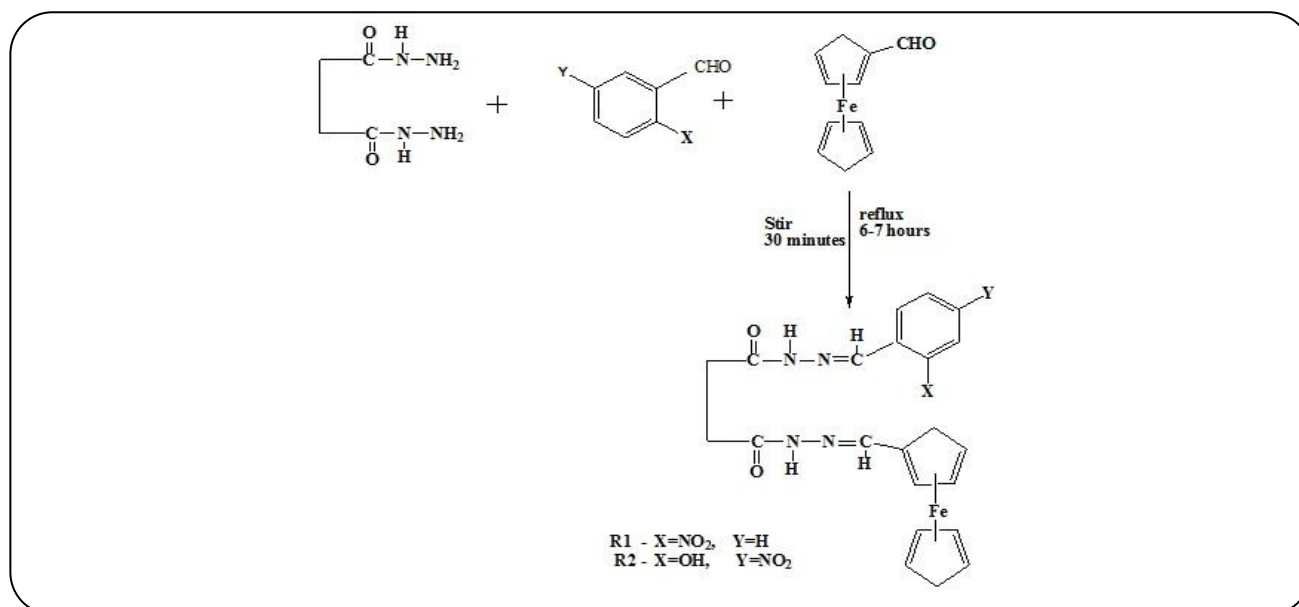
$NiCl_2$ ,  $Cd(OAc)_2$ ,  $CuCl_2$  and  $MnCl_2$ , used in the cyclic voltammetry and UV-Visible titration studies were procured from Sigma–Aldrich. HPLC grade acetonitrile acquired from E- Merck and spectral grade absolute ethanol secured from Commercial Alcohols, Canada was used for spectral studies. Pure tetrabutylammoniumperchlorate [99+%] obtained from Chemical Center, Mumbai was used as such without purification.

### Instruments

Mass spectra were documented in Bruker Daltonics esquire 3000 spectrometers. Proton NMR spectra were taken in BRUKER AVANCE spectrometer [500 MHz] engaging  $C_2D_5OD$  solvent. Using KBr pellets, FTIR spectra in the range of  $400-4000\text{ cm}^{-1}$  were recorded in Perkin-Elmer 337 spectrometer. SHIMADZU MODEL UV-1800 240 V spectrophotometer was tied up to register UV–visible spectral studies between 200 and 800 nm. Electrochemical analyzer 1200B model having glassy carbon as the working electrode, Ag/AgCl as the reference electrode, and platinum as the counter electrode was employed to register cyclic voltammograms. TBAP solution (handled under care) was used as a supporting electrolyte. Elements like C, H, and N were analyzed with Herarus C-H-N rapid analyzer.

### Synthesis of *N*'1-((*E*)-2-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide [R1]

A modified procedure reported in the literature [23] was used to prepare the precursor compound succinic acid dihydrazide. To a solution of succinic anhydride (0.05 moles in 50 mL of ethanol) another solution containing hydrazine hydrate (0.12 moles) in 150 mL of ethanol was added in drops along with stirring and then the mixture was refluxed for 24 hours. Precipitated succinic acid dihydrazide was dried and recrystallized from ethanol. To a clear solution (0.01 mol) of purified succinic acid dihydrazide in 50 mL of ethanol, a mixture of 2-nitrobenzaldehyde (0.01 mol) and ferrocenecarboxaldehyde (0.01 mol) in 150 mL ethanol was added. After half an hour of stirring, the reaction mixture was refluxed for 6-7 hours. The progress of the reaction was checked at various time intervals using a thin-layer chromatographic technique. The cooled reaction mixture was filtered and the filtrate was concentrated to get reddish yellow colored *N*'1-((*E*)-2-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide. Crude sample



Scheme 1: Scheme of synthesis

was purified in a silica gel column using ethanol as eluent. Color: Dark reddish orange. Yield: 0.5332 g (91%), m.p. 180 °C.

#### Synthesis of *N*'1-((*E*)-2-hydroxy-5-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide [R2]

Succinic acid dihydrazide solution (0.01 mole in 50 mL of ethanol) was added with stirring to another solution having 0.01 mole of 2-hydroxy-5-nitrobenzaldehyde and 0.01 mole of ferrocenecarboxaldehyde in 150 mL ethanol. The above mixture was stirred for half an hour and then refluxed for 6-7 hours. The thin-layer chromatographic technique was used to check the progress of the reaction at various time intervals. After cooling, the reaction mixture was filtered. The red colored solid was obtained after concentrating the filtrate. Purification of the crude product was carried out in a column having silica gel as the stationary phase and ethanol as eluent. Color: reddish orange, Yield: 0.6015 g, (91%), m.p. 182 °C.

#### Binding energy calculation using molecular docking

Investigation of the binding manner of the manufactured compounds R1 and R2 with the target protein available in the microorganisms was done in Auto Dock version 1.5.6 [25] running on Windows 7. Target proteins of the enzymes were extracted from RCSB, Protein Data Bank (PDB). MGL tools of Auto Dock were employed to get the docking score. ChemSketch was

engaged to draw the structures of the compounds R1 and R2 and converted them to 3D structures by utilizing 3D optimization tool. Ligands were geometrically optimized using a ligand module. Force field MMFF94 was engaged to compute the value of partial atomic charges. Selected proteins were docked with synthesized ligands using Auto Dock tools. H-bond interactions and binding affinity were analyzed for the best-docked pose obtained from conformation.

#### Assessment of antimicrobial activity

Experiments in triplicate were carried out using standard procedure [24] against two gram-positive bacteria, two gram-negative bacteria, and two fungi at appropriate temperatures. The average value arrived from the three experiments was considered for analysis.

#### Analysis of elements

Experimental data obtained on elemental analysis of the synthesized compounds match with the molecular formula values. R1 (Found (%): C, 55.65; H, 4.39; N, 14.70; Fe, 11.41; Calc. for C<sub>22</sub>H<sub>21</sub>N<sub>5</sub>O<sub>4</sub>Fe: C, 55.60; H, 4.43; N, 14.76; Fe, 11.45). R2 (Found (%): C, 55.36; H, 4.10; N, 14.73; Fe, 11.40; Calc. for C<sub>22</sub>H<sub>21</sub>N<sub>5</sub>O<sub>5</sub>Fe: C, 55.30; H, 4.43; N, 14.76; Fe, 11.45). The presence of molecular peak (ESI) m/z at 474 and 482 respectively for the compounds *N*'1-((*E*)-2-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide and *N*'1-((*E*)-2-hydroxy-5-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)

succinohydrazide on mass spectral analysis, confirm the formation of expected receptors.

### FT-IR Spectral analysis

The FT-IR spectral peak of compound R1 observed around  $487\text{ cm}^{-1}$  is related to the tilt stretching vibration of the ferrocene cyclopentadienyl ring and the  $829\text{ cm}^{-1}$  peak corresponds to C-H out-of-plane bend vibration. The -C-C-H bending vibration in the cyclopentadienyl ring [26] emerged between  $935\text{ cm}^{-1}$  to  $1244\text{ cm}^{-1}$ . The peaks for symmetric stretching vibrations of  $-\text{NO}_2$ ,  $-\text{CH}=\text{N}$  (imine) stretching vibration, and amide  $-\text{C}=\text{O}$  stretching arose at  $1344\text{ cm}^{-1}$ ,  $1523\text{ cm}^{-1}$  and  $1673\text{ cm}^{-1}$  respectively [27]. The stretching vibration of a secondary amine and water of hydration appeared as a single peak at  $3199\text{ cm}^{-1}$ . The FT-IR spectral behavior of compound R2 is similar to that of R1 except for the intensity of  $3199\text{ cm}^{-1}$  peak wherein the phenolic -OH group stretching vibration also merges with the stretching vibration of secondary amine and water of hydration.

### NMR Spectral analysis

Nuclear magnetic resonance spectrum of R1 in  $\text{C}_2\text{D}_5\text{OD}$  solvent contains appropriate peaks and are assigned accordingly  $\delta$ , (ppm); 8.4 (s, 2H, NCH), 7.5 (m, 4H, aromatic), 4.7 (m, 2H, cp subst), 4.2 (m, 2H, cp subst), 4.1 (s, 5H, cp unsubst), 2.7 (2s, 4H,  $2\text{CH}_2$ ), 1.2 (s, 2H, 2NH) (Fig. 1) for R2  $\delta$ , (ppm); 8.5 (s, 2H, NCH), 7-7.4 (m, 4H, aromatic), 4.5 (m, 2H, cp subst), 4.3 (m, 2H, cp subst), 4.1 (s, 5H, cp unsubst), 2.7 (2s, 4H,  $2\text{CH}_2$ ), 1.3 (s, 2H, 2NH).

## RESULTS AND DISCUSSION

### Examination of sensing behavior of R1 & R2

Compound R1 in acetonitrile exhibits two peaks around 249 nm & 281 nm and a broad peak near 469 nm (Fig. 2a). In alcohol, R1 displays two peaks near 251 nm and 289 nm (Fig. 2b). Aromatic ring  $\pi-\pi^*$  transition is assigned for the peaks in the ultraviolet region [28] and d-d transition is earmarked for visible region peak [29].

UV-Visible titration method was engaged to find out the incarceration ability of receptors with various metal ions. About 2.5 mL of receptor solution ( $10^{-5}\text{ M}$ ) was taken in the quartz cell and 20  $\mu\text{L}$  aliquots of metal solutions ( $10^{-2}\text{ M}$ ) were added. Absorbance spectra were recorded for metal ion-free and metal ion-added sensor solutions.

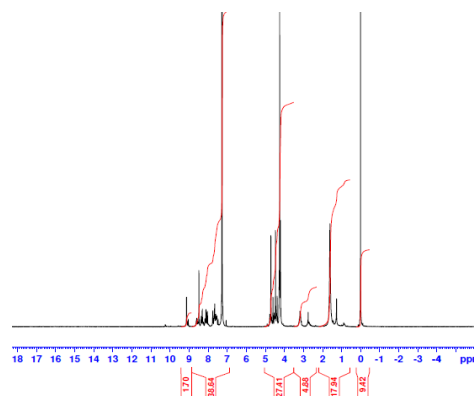


Fig.1: Proton NMR spectrum of sensor R1.

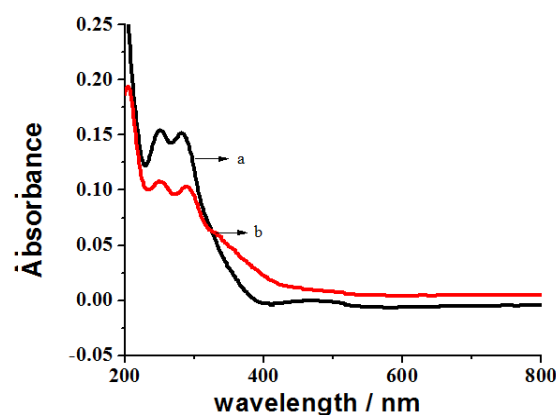


Fig. 2: Absorbance spectrum of R1 in a) acetonitrile b) ethanol.

Based on the solubility of selected metal salt for investigation, either the solution of receptor in acetonitrile or ethanol was utilized to chronicle the electronic spectra. Development of new peaks around 301 nm, 352 nm, and 457 nm for the addition of  $\text{Cu}^{2+}$  ions with the receptor at the expense of basic peaks of R1 exposes the sensing aptitude (Fig. 3a). The generation of 457 nm peak (Fig. 3b) which is responsible for the metal-to-ligand charge transfer [30] that has acquired after the coordination of  $\text{Cu}^{2+}$  ions with receptor also ascertains the sensing ability.

Successive addition of  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ , and  $\text{Pb}^{2+}$  ions generate new peaks around 237, 246, and 238 nm respectively (Fig. 4) with simultaneous disappearance of peaks responsible for receptor. The cumulative addition of  $\text{Mn}^{2+}$  and  $\text{Cd}^{2+}$  ions resulted in an increase in absorbance value along all wavelength regions [28]. Above findings advocate that R1 is capable of sensing  $\text{Cu}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Mn}^{2+}$  and  $\text{Cd}^{2+}$  ions.

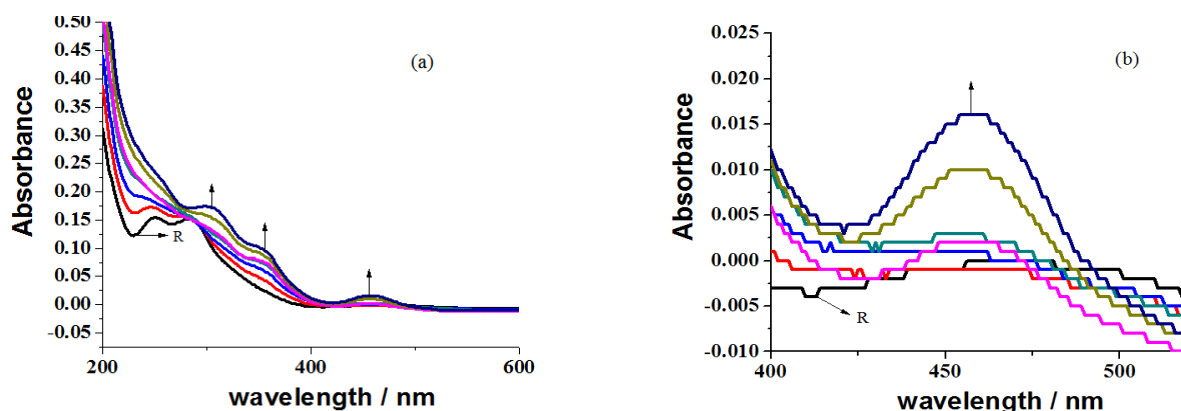


Fig. 3: a) Spectral changes noticed for the addition of  $\text{Cu}^{2+}$  ions to R1 b) Generation of MLCT band.

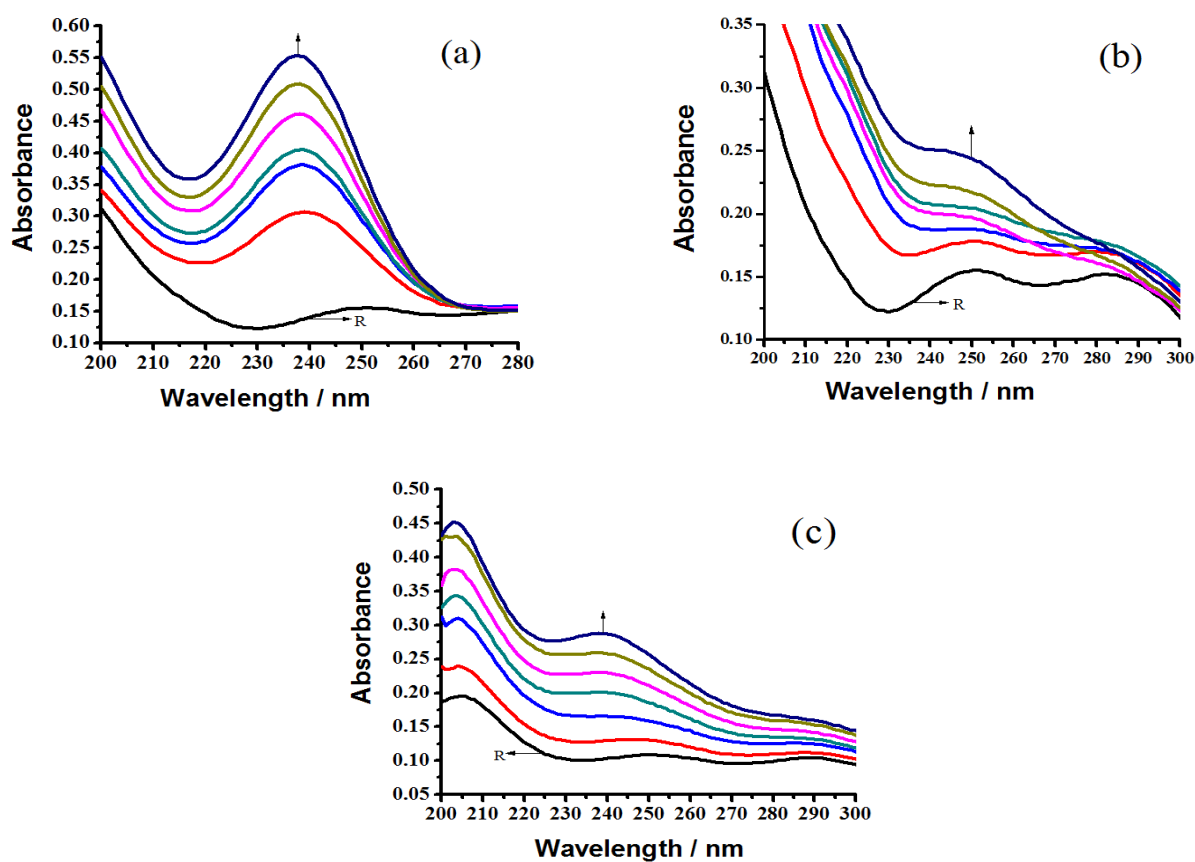


Fig.4: Development of a new peak in the spectrum of R1 for the addition of a)  $\text{Hg}^{2+}$  ions b)  $\text{Ni}^{2+}$  ions c)  $\text{Pb}^{2+}$  ions.

A shoulder around 282 nm in acetonitrile and a broad peak at 346 nm in ethanol solvents were recorded for the  $\pi$ - $\pi^*$  transition of compound R2. Formation of new peaks near 253 nm, 311 nm, and 461 nm (Fig. 5a, b, c) with the disappearance of 282 nm shoulder of the receptor for the successive addition of  $\text{Cu}^{2+}$  ions to R2 reveals the sensing ability of the receptor. Earlier report [30] had assigned

metal-ligand charge transition for the appearance of a new peak around 460 nm.

The binding ability of R2 towards  $\text{Hg}^{2+}$ ,  $\text{Mn}^{2+}$ , and  $\text{Cd}^{2+}$  ions is exposed by the formation of a new peak at 240 nm for  $\text{Hg}^{2+}$  (Fig. 6a), conversion of broad peak to prominent peak around 346 nm for  $\text{Mn}^{2+}$  (Fig. 6b) and  $\text{Cd}^{2+}$  (Fig. 6c) respectively. The disappearance of the shoulder peak of

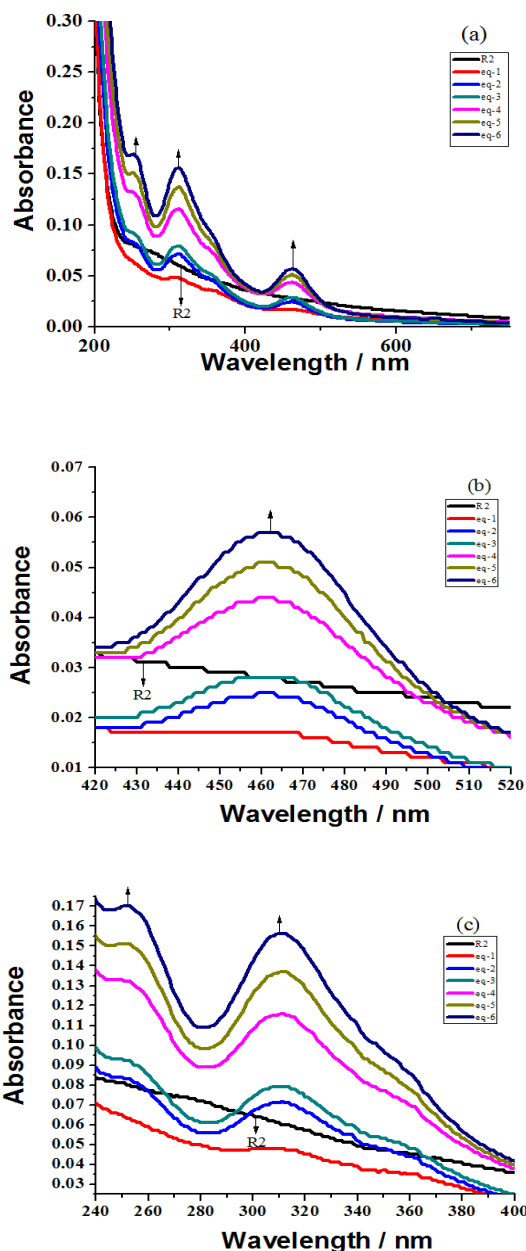


Fig.5: Spectral changes of R2 for the addition of  $\text{Cu}^{2+}$  ion a) Overall changes b) Generation of MLCT band c) Development of new peaks.

the receptor with increased absorbance value in all wavelength regions for the addition of  $\text{Pb}^{2+}$  and  $\text{Ni}^{2+}$  ions shows the sensing ability of R2 towards these metal ions.

#### Electrochemical investigation of sensing studies

Analysis of the voltammograms of R1 (Fig. 7) recorded with different scan rates for  $1 \times 10^{-3}$  M solution containing oxidation peak at 0.712 V and reduction peak around 0.566 V.

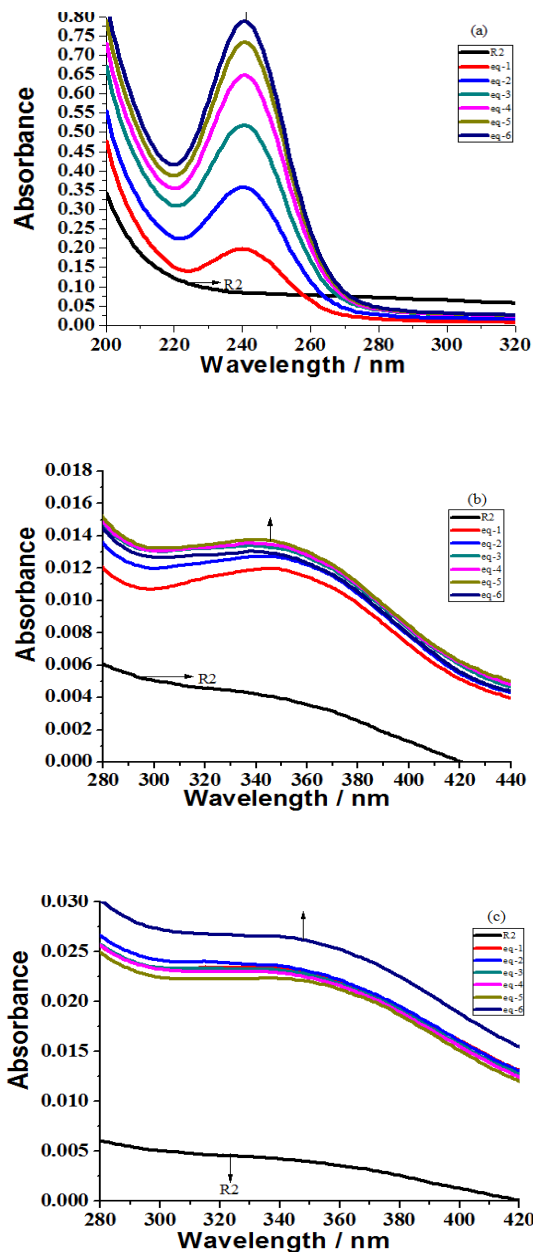


Fig.6: Appearance of a new peak in the spectrum of R2 for the addition of a)  $\text{Hg}^{2+}$  b)  $\text{Mn}^{2+}$  c)  $\text{Cd}^{2+}$ .

Further increasing trend in  $\Delta E_p$ ,  $I_{pa}$  &  $I_{pc}$  values (Table 1) with an increase in scan rate is observed. Overblown  $\Delta E_p$  values (146-161 mV, than the anticipated 59 mV) noticed for all scan rates suggest a quasi-reversible reduction of  $\text{Fe}^{2+}$  present in ferrocene moiety [31].

CV- titration studies were carried out by adding 20  $\mu\text{L}$  (using micropipette) of either  $1 \times 10^{-3}$  M (Fig. 8a) or  $1 \times 10^{-1}$  M (Fig. 8b) metal salts solution to 10 mL of  $1 \times 10^{-3}$  M R1

Table 1: Electrochemical parameters for RI.

Scan Rate- mV/ sec	$E_{pa}$ (V)	$E_{pc}$ (V)	$\Delta E_p$ (V)	$E_{1/2}$ (V)	$I_{pa} \times 10^{-6}$ ( $\mu A$ )	$I_{pc} \times 10^{-6}$ ( $\mu A$ )
Solvent - Acetonitrile						
20	0.712	0.566	0.146	0.639	-2.907	1.779
50	0.721	0.566	0.155	0.643	-4.590	3.260
100	0.719	0.564	0.155	0.641	-6.411	4.978
Solvent - Ethanol						
20	0.712	0.551	0.161	0.631	-2.564	1.311
50	0.708	0.559	0.149	0.633	-3.811	2.625
100	0.706	0.555	0.151	0.630	-5.327	4.245

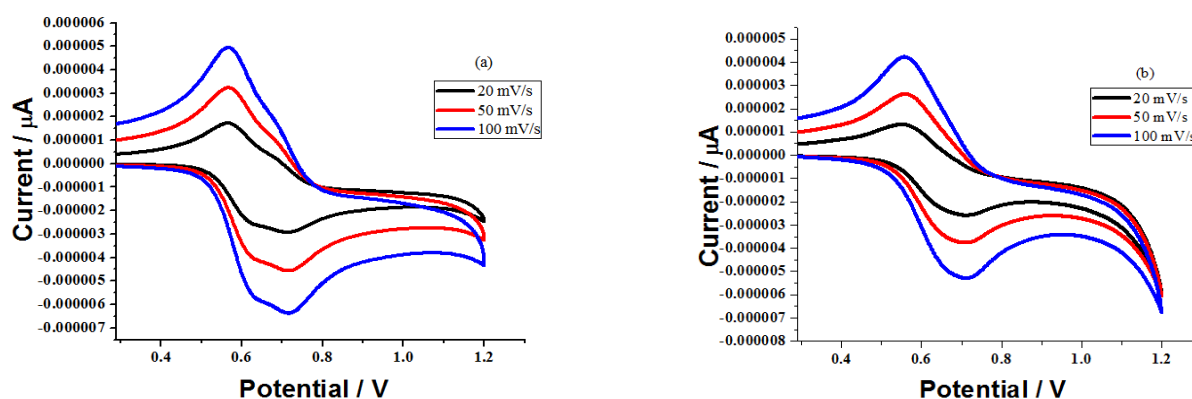
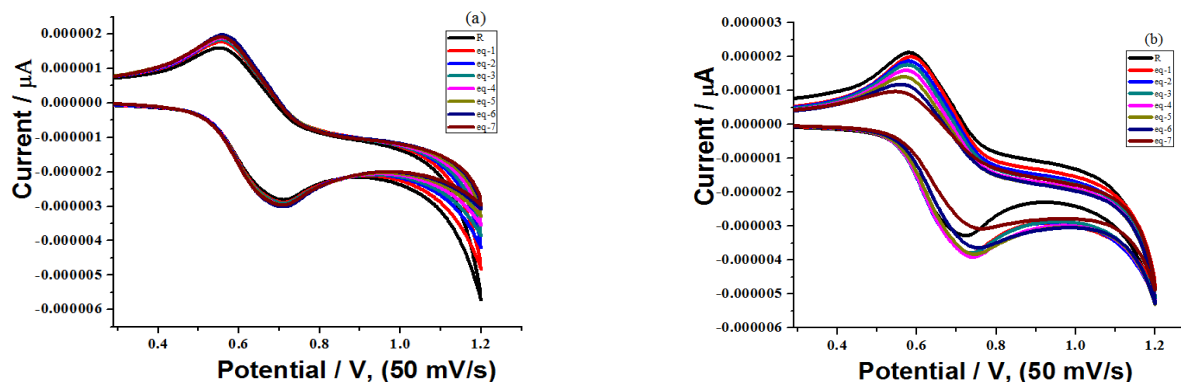


Fig.7: Cyclic voltammogram of RI recorded with different scan rate a) acetonitrile b) ethanol.

Fig.8: Changes in voltammogram of RI for the addition of  $Cd^{2+}$  ions a)  $1 \times 10^{-3} M$  b)  $1 \times 10^{-1} M$ .

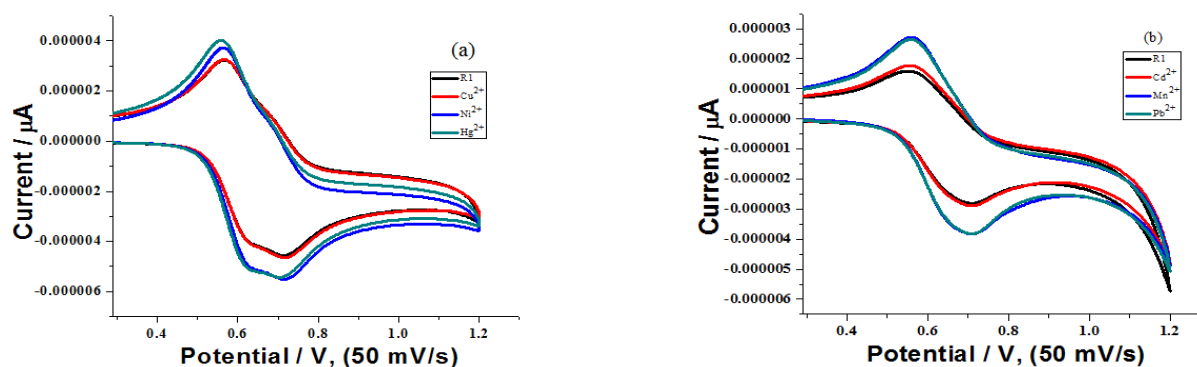
a solution taken in the three-compartment cell up to 7 equivalents and cyclic voltammograms (scan rate - 50 mV/s) were recorded for each addition. Changes in voltammograms contain positive potential shifts for the oxidation peak and negative potential shifts for the reduction peak which support the interaction operating

between the receptor and added metal ions [32]. As an example, the voltammograms registered for the addition of  $Cd^{2+}$  ions are presented in Fig. 8.

A varied amount of  $I_{pa}$  values observed (Table 2) for the addition of dissimilar metal ions (Fig. 9) having  $1 \times 10^{-3} M$  concentration exposes the difference in binding aptitude of

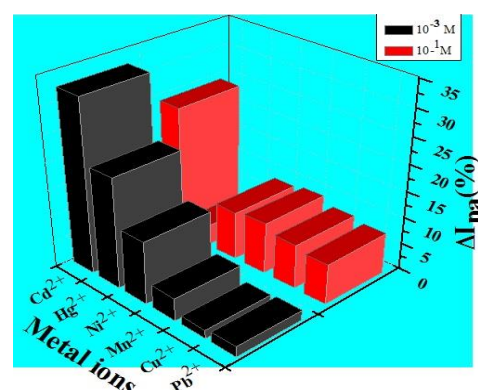
**Table 2: Electrochemical data for the addition of various metal ion solution (R1,  $10^{-3}$  M /  $M^{2+}$ ,  $10^{-3}$  M) (Scan Rate - 50 mV/s).**

Addition	$E_{pa}$ (V)	$E_{pc}$ (V)	$\Delta E_p$ (V)	$E_{1/2}$ (V)	$I_{pa} \times 10^{-6}$ ( $\mu A$ )	$I_{pc} \times 10^{-6}$ ( $\mu A$ )
Solvent - Acetonitrile						
Receptor	0.721	0.566	0.155	0.643	-4.590	3.260
$Cu^{2+}$	0.719	0.566	0.153	0.637	-4.652	3.314
$Ni^{2+}$	0.721	0.564	0.157	0.642	-5.553	3.704
$Hg^{2+}$	0.706	0.558	0.148	0.632	-5.434	4.032
Solvent - Ethanol						
Receptor	0.708	0.559	0.149	0.633	-3.811	2.625
$Cd^{2+}$	0.710	0.553	0.157	0.631	-2.898	1.769
$Mn^{2+}$	0.708	0.559	0.149	0.633	-3.846	2.743
$Pb^{2+}$	0.702	0.559	0.143	0.630	-3.846	2.620

**Fig. 9: Cyclic voltammograms recorded (scan rate - 50 mV/s) for the addition of different metal ion to R1 a) acetonitrile b) ethanol.**

metal cations. Apart from that, it is a measure of repulsive force existing between the oxidized ferrocene moiety and coordinated metal cation [33]. The percentage difference in  $I_{pa}$  values ( $\Delta I_{pa}$  %) calculated for the oxidation current value of receptor solution and metal ions added to receptor solution reflects the sensing order of R1 as  $Cd$  (32.6) >  $Hg$  (20.8) >  $Ni$  (11.9) >  $Mn$  (4.3) >  $Cu$  (1.6) >  $Pb$  (1.9).

The  $\Delta I_{pa}$  (%) values (Table 3) calculated from the noted  $I_{pa}$  values for the addition of  $1 \times 10^{-1}$  M metal salt solutions to  $1 \times 10^{-3}$  M R1 solution disclose the sensing power of R1 as  $Cd^{2+}$  (24.3) >  $Mn^{2+}$  (9.7) >  $Ni^{2+}$  (9.1) >  $Cu^{2+}$  (8.4) >  $Pb^{2+}$  (7.6) >  $Hg^{2+}$  (3.8). Careful analysis of the  $\Delta I_{pa}$  (%) values estimated for the addition of  $1 \times 10^{-3}$  M and  $1 \times 10^{-1}$  M concentrations of various metal ions convey that R1 is potent towards Cd, Hg & Ni at lower concentrations and persuasive to Cd, Mn & Ni at higher concentration (Fig. 10).

**Fig.10: Comparison chart for sensing ability of R1.**

The inclination of voltammograms perceived for R2 with different scan rates is similar to R1 with an increase in value of  $\Delta E_p$ ,  $I_{pa}$ , and  $I_{pc}$  (Table 4).

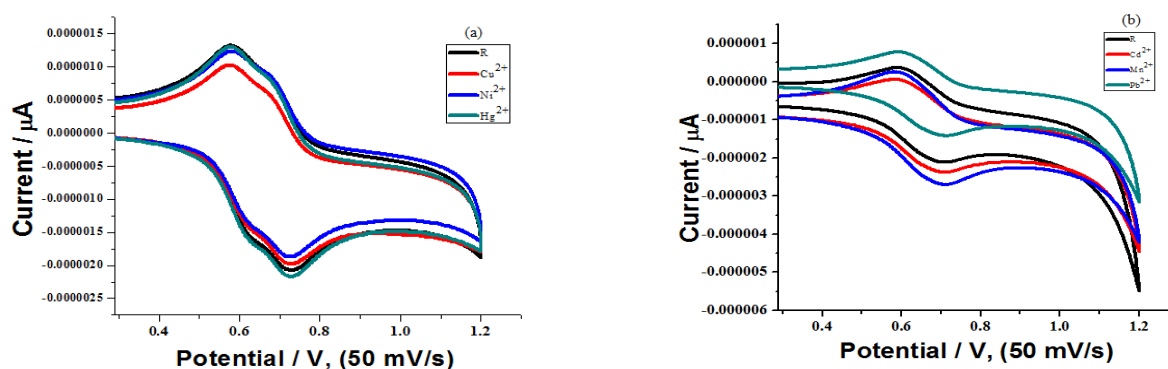


**Table 3: CV data obtained (scan rate- 50 mV/s) for the addition of  $1 \times 10^{-1} M$  metal salt solutions to  $1 \times 10^{-3} M$  R1.**

Addition	$E_{pa}$ (V)	$E_{pc}$ (V)	$\Delta E_p$ (V)	$E_{1/2}$ (V)	$I_{pa} \times 10^{-6}$ ( $\mu A$ )	$I_{pc} \times 10^{-6}$ ( $\mu A$ )
Solvent - Acetonitrile						
Receptor	0.721	0.566	0.155	0.643	-4.590	3.260
$Cu^{2+}$	0.723	0.595	0.128	0.659	-4.684	2.983
$Ni^{2+}$	0.725	0.595	0.13	0.66	-5.821	3.585
$Hg^{2+}$	0.739	0.603	0.136	0.671	-4.413	3.134
Solvent - Ethanol						
Receptor	0.708	0.559	0.149	0.633	-3.811	2.625
$Cd^{2+}$	0.729	0.584	0.145	0.656	-3.806	1.987
$Mn^{2+}$	0.715	0.582	0.407	0.648	-3.338	2.370
$Pb^{2+}$	0.719	0.586	0.133	0.652	-3.477	2.425

**Table 4: Electrochemical parameters for R2 with different scan rate.**

Scan Rate mV/s	$E_{pa}$ (V)	$E_{pc}$ (V)	$\Delta E_p$ (V)	$E_{1/2}$ (V)	$I_{pa} \times 10^{-6}$ ( $\mu A$ )	$I_{pc} \times 10^{-6}$ ( $\mu A$ )
Solvent - Acetonitrile						
20	0.725	0.576	0.149	0.650	-1.227	0.531
50	0.727	0.576	0.151	0.648	-1.969	1.079
100	0.723	0.581	0.142	0.652	-2.846	1.806
Solvent - Ethanol						
20	0.694	0.607	0.087	0.650	-1.779	-1.853
50	0.694	0.593	0.101	0.643	-2.096	3.530
100	0.701	0.593	0.108	0.647	-2.745	1.118

**Fig.11: Voltammograms (scan rate- 50 mV/s) recorded for the addition of different metal ions to R2 a) acetonitrile b) ethanol.**

Employing the same procedure utilized for R1, CV titration studies (scan rate - 50 mV/s) were carried out for R2 with various metal salts solutions (Fig.11) and the difference in  $I_{pa}$  amount noticed (Table 5) exposes the

change in the ability of the receptor to bind with the different metal ion. Binding order priority based on the  $\Delta I_{pa}$  (%) under equimolar addition (Table 5) is  $Pb^{2+}$  (54.9%) >  $Cd^{2+}$  (33.1%) >  $Mn^{2+}$  (21.5%) >  $Hg^{2+}$  (17.5%) >  $Ni^{2+}$  (12.3%) >  $Cu^{2+}$  (4.7%)

**Table 5: CV titration data (scan rate- 50 mV/s) for the addition of  $1 \times 10^{-3}$  M metal salt solution to  $1 \times 10^{-3}$  M R2.**

Addition	$E_{pa}$ (V)	$E_{pc}$ (V)	$\Delta E_p$ (V)	$E_{1/2}$ (V)	$I_{pa} \times 10^{-6}$ ( $\mu A$ )	$I_{pc} \times 10^{-6}$ ( $\mu A$ )
Solvent - Acetonitrile						
Receptor	0.727	0.576	0.151	0.648	-1.969	1.079
$Cu^{2+}$	0.729	0.574	0.155	0.651	-1.993	1.028
$Ni^{2+}$	0.723	0.580	0.143	0.651	-1.859	1.230
$Hg^{2+}$	0.727	0.578	0.149	0.652	-2.185	1.309
Solvent - Ethanol						
Receptor	0.694	0.593	0.101	0.643	-2.096	3.530
$Cd^{2+}$	0.704	0.586	0.121	0.645	-2.362	5.278
$Mn^{2+}$	0.702	0.586	0.116	0.644	-2.718	2.771
$Pb^{2+}$	0.704	0.595	0.109	0.649	-1.425	7.829

**Table 6: CV titration data (scan rate- 50 mV/s) for the addition of  $1 \times 10^{-3}$  M metal salt solution to  $1 \times 10^{-1}$  M R2.**

Addition	$E_{pa}$ (V)	$E_{pc}$ (V)	$\Delta E_p$ (V)	$E_{1/2}$ (V)	$I_{pa} \times 10^{-6}$ ( $\mu A$ )	$I_{pc} \times 10^{-6}$ ( $\mu A$ )
Solvent - Acetonitrile						
Receptor	0.727	0.576	0.151	0.648	-1.969	1.079
$Cu^{2+}$	0.715	0.599	0.116	0.657	-2.978	1.830
$Ni^{2+}$	0.772	0.634	0.138	0.703	-2.390	1.341
$Hg^{2+}$	0.783	0.640	0.143	0.711	-1.901	1.270
Solvent - Ethanol						
Receptor	0.694	0.593	0.101	0.643	-2.096	3.530
$Cd^{2+}$	0.762	0.659	0.103	0.710	-1.469	5.483
$Mn^{2+}$	0.696	0.605	0.091	0.650	-1.235	6.451
$Pb^{2+}$	0.770	0.663	0.107	0.716	-1.661	9.064

and multimolar addition (Table 6) is  $Pb^{2+}$  (61) >  $Mn^{2+}$  (45) >  $Cu^{2+}$  (41) >  $Cd^{2+}$  (35%) >  $Hg^{2+}$  (15) >  $Ni^{2+}$  (14.3). Fig. 12 symbolizes the comparison between the sensing ability of R2 and the moderate metal concentration of metal ions required.

#### Antimicrobial Studies

Antibacterial activity of R1 & R2 was carried out by disc diffusion method, having Mueller Hinton Agar base, against *Streptococcus faecalis*, *Staphylococcus aureuse*, *Salmonella typhimurium* and *Escherichia coli* (Fig. 13) and anti-fungal studies were conducted with Sabouraud's Dextrose agar as a base against fungi *Candida albicans*

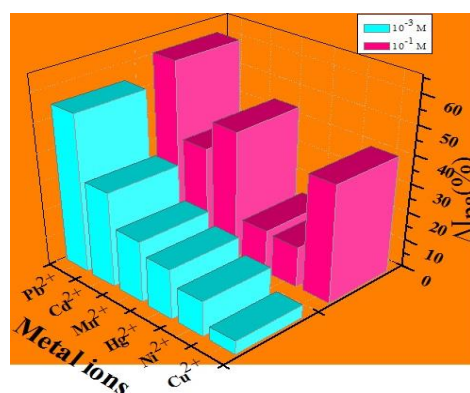
**Fig. 12: Comparison chart for sensing ability of R2.**

Table 7: In-vitro antimicrobial studies data.

S.No	Microorganisms	Control	R1	R2	Ciprofloxacin/Ketoconazole
zone of inhibition in mm for bacteria					
1	Staphylococcus aureus	-	12	06	25
2	Streptococcus faecalis	-	-	-	24
3	Escherichia coli	-	09	07	12
4	salmonella typhimurium	-	10	08	27
zone of inhibition in mm for fungi					
1	Candida albicans	-	-	06	06
2	Aspergillus niger	-	12	10	25

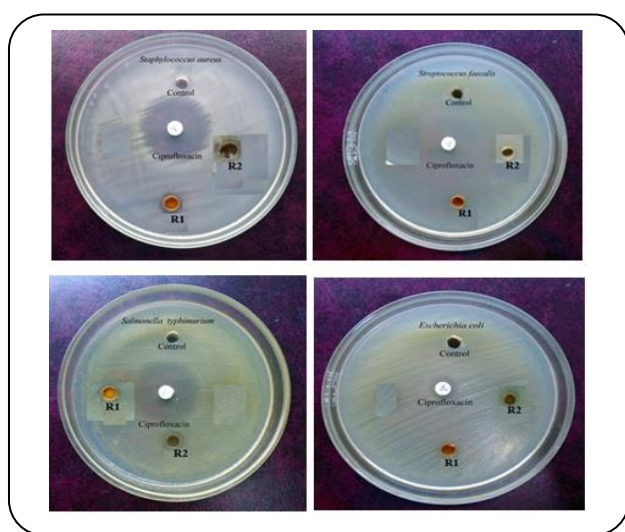


Fig. 13: Zones of inhibition for a) SA b) SF c) ST d) EC.



Fig.14: Zones of inhibition for a) CA b) AN.

and *Aspergillus niger* (Fig. 14). Data of growth inhibition distance perceived for R1 & R2 in the antimicrobial analysis are presented in Table 7.

Compound R2 prohibition efficiency against fungi *Candida albicans* is 100% in comparison with the standard drug ketoconazole, which is unusual and demands more focused research by the pharmacist to develop a new

formulation of antifungal agent [22]. The resistivity of R1 and R2 to resist the growth of fungi *aspergillus niger* is also comes to around 50%. The retardant nature of R1 & R2 towards bacteria is on par with standard ciprofloxacin, except for *Streptococcus faecalis*.

#### Molecular docking studies

Molecular docking studies were carried out using software, mentioned in the experimental section, for the proteins of the microorganisms as stated here: 6TZ6- *Candida albicans*, 3K4Q- *Aspergillus niger*, 7BU2- *Escherichia coli*, 4YXB- *salmonella typhimurium*, 6KVQ- *Staphylococcus aureus* and 1PTF- *Streptococcus faecalis* and the results arrived are presented in (Table 8) The binding posture of the ligand and proteins in 3D and 2D angle are given in figure -15. Fungi proteins 3K4Q & 6TZ6 bind comfortably with both the ligands by giving binding energy value greater than -5 Kcal mol<sup>-1</sup> which exposes the higher antifungal activity of newly synthesized receptors [34]. In addition to that compound R1 & R2 displayed better scores against bacteria proteins 4YXB & 7BU2.

#### CONCLUSIONS

Unsymmetrical Schiff base compounds comprising of ferrocene occupied azomethine group at one side and aromatic aldehyde attached mine on the other side is prepared in a single step reaction. The newly synthesized materials *N*'1-((*E*)-2-(nitro)benzylidene)-*N*'4-((*E*)-2(ferrocenylidene)succinohydrazide and *N*'1-((*E*)-2-hydroxy-5-(nitro)benzylidene)-*N*'4-((*E*)-2-(ferrocenylidene)succinohydrazide were upheld by FT-IR, <sup>1</sup>HNMR and Mass spectrum analysis. Ability of the

Table 8: Data obtained in molecular docking studies.

PDB	Free binding energy, kcal mol <sup>-1</sup>		R1		R2	
	R1	R2	Formation of hydrogen bonds with receptor and amino acids	Distance(Å)	Formation of hydrogen bonds with receptor and amino acids	Distance(Å)
1PTF	-4.04	-3.09	33-ILE 45-LYS 43-ASN	3.24 3.18 3.39	82-GLN 78-VAL	3.64 3.38
3K4Q	-5.16	-5.79	49-ARG 123-ARG 124-TYR	3.18 3.28 3.31	271-ASP 272-TYR 366-ILE	3.21 3.86 3.96
4YXB	-4.55	-7.57	25-PHE 27-ASP 45-PRO	3.54 3.17 3.27	25-PHE 27-ASP 47-GLU	3.42 3.27 3.12
6KVQ	-3.14	-3.98	116-VAL 115-PRO 119-LYS	2.98 3.38 3.62	25-ASN 139-GLU 179-MET 183-ALA	3.85 3.28 3.31 3.96
6TZ6	-6.29	-6.33	58-GLN 97-TYR	3.40 3.82	40-PHE 46-ARG 59-VAL 97-TYR	2.78 3.86 3.75 3.85
7BU2	-5.42	-7.81	264-VAL 284-GLY	3.96 3.47	261-VAL 263-ALA 332-ARG	3.47 3.76 3.18

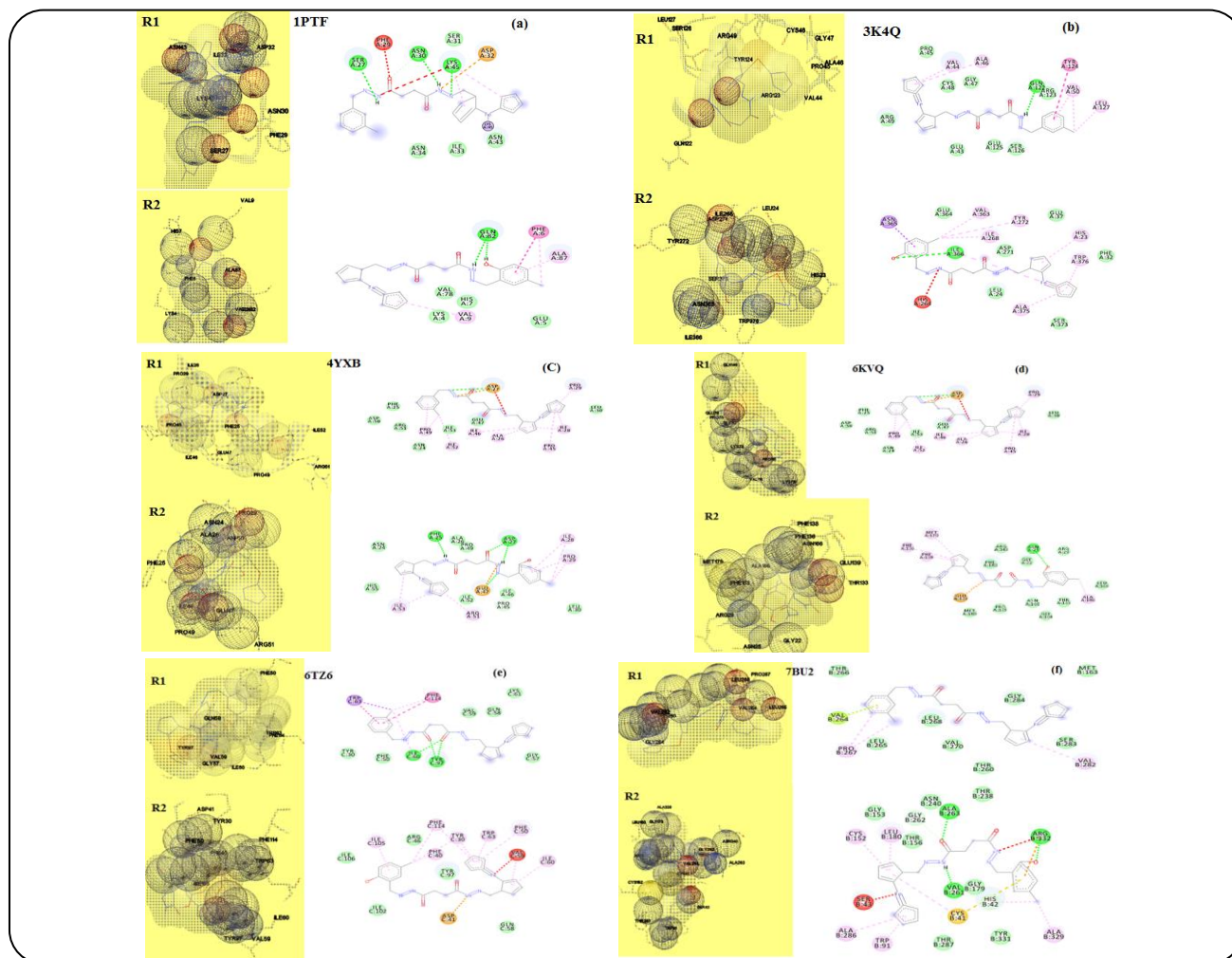


Fig.15: 3D and 2D view binding of R1 and R2 with a) 1PTF b) 3K4Q c) 4YXB d) 6KVQ e) 6TZ6 f) 7BU2.

receptor to sense  $\text{Hg}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Ni}^{2+}$  and  $\text{Cu}^{2+}$  ions is identified in UV-Visible titration method. The effective concentration of metal ions required for a particular concentration of sensor solution is arrived at from the percentage amount of  $\Delta I_{pa}$  values calculated from the anodic current values ( $I_{pa}$ ) noticed from the cyclic voltammograms recorded for the receptor solution containing different metal ions. Hundred percent efficiency, to prevent the growth of the fungus *Candida albicans* by compound R2, is recognized from *in-vitro* and molecular docking studies which entice pharmaceutical research to be undertaken.

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