Chemoenzymatic Enantioselective Formal Synthesis of (-)-Gephyrotoxin-223

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ABSTRACT: (-)-Gephyrotoxin-223 was formally synthesized from chiral synthon 1 which has been chemoenzymatically synthesized in the presence of Candida Antartica lipase.

KEY WORDS: Gephyrotoxin, Indolizidine, Chemoenzymatic, Candida Antartica Lipse, Alkaloids, Desymmetrization.

INTRODUCTION
For a wide range of simple bicyclic alkaloids that have been found in skin extracts of dendrobatid frogs and mantelline frogs, the term “izidine” alkaloids might be used [1]. The “izidine” alkaloids detected in amphibian skin include 3,5-disubstituted pyrrolizidines, 3,5-disubstituted and 5-monosubstituted indolizidines, 5,8-disubstituted indolizidines, 5,6,8-trisubstituted indolizidines. Many of them probably originate from dietary ants. In this paper, we will present our work on the synthesis of (-)-gephyrotoxin-223 as one of the 3,5-disubstituted indolizidines.

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In 1978, 3,5-disubstituted indolizidine structures were proposed [2] for three alkaloids found in the dendrobatid frog *Dendrobates histrionicus*. One of them was (-)-gephyrotoxin-223 (indolizidine 223AB) of which the 5E, 9E isomer has been found in a Colombian frog *Dendrobates histrionicus* and its 5Z,9Z isomer was the sole diastereomer found in Panamanian *Dendrobates specious*.

About 15 alkaloids detected in extracts of amphibian skin have been proven to be 3,5-disubstitued indolizidines and they represent another alkaloid class present as venom constituents in myrmicine ants. Recently, both monomorine I and the amphibian diastereomer 5E,9E–195B were found in a Puerto Rican myrmicine ant.

When Dendrobatid frogs (*Dendrobates auratus*), were fed Pharoah’s ants (*Monomorium pharaonis*), they efficiently accumulated the ant alkaloid monomorine I and two minor 3,5-disubstitued indolizidines in their skin.

Indolizidine 5E,9E–195B was a major alkaloid in a dendrobatid frog (*Dendrobates auratus*) raised in outside cages in Hawaii.

It seems highly likely that the 3,5-disubstitued indolizidines found in amphibian skin are the result of sequestration from myrmicine ants.

Of the 5-monosubstituted indolizidines, we have reported [3,4] enantioselective synthesis of the enantiomeric pairs of indolizidine 167B and indolizidine 209D and of the 3,5-disubstituted indolizidines, we have performed the formal asymmetric synthesis of (-)-gephyrotoxin-223 (indolizidine 223AB) via enzymatic desymmetrization.

**Retrosynthetic analysis**

Our construction strategy for 3,5-disubstituted indolizidines is based on the retrosynthetic analysis illustrated in scheme 1. This analysis introduces the chiral synthon 1 as the key intermediate which was prepared.
from 2,6-dicarboxylic acid 4 in several steps. As we know, the enzymatic hydrolysis of meso-diester 2 in the presence of Aspergillus niger lipase (ANL) \([5,6]\) provided the corresponding (+)-(2R,6S)-monoacetate 1, while the enzymatic acetylation of meso-diol \([3,4]\) 3 gave the (-)-(2S,6R)-monoacetate 1. These two enantiomeric pairs were used as chiral building blocks in the enantioselective synthesis of both enantiomers of 3,5-disubstituted indolizidines.

**DISCUSSION**

We report here the chemoenzymatic enantioselective formal synthesis of both enantiomers of (-)-gephyrotoxin-223 (indolizidine \(223\text{AB}\)) from the unstable aldehyde 11, readily prepared \([3]\) from the (2R,6S)-monoacetate 1 as chiral synthon which was prepared as shown in scheme 2, 3, 4. The aldehyde 11 was added to Wittig ylide, ethyl (triphenylphosphoranylidene)acetate, to provide the olefin 12, which was hydrogenated over Pearlman’s Catalyst to yield aminoester 13. The protection of the amino group of 13 was effected using benzylchloroformate giving the desired intermediate 14 \([\alpha]_D^25 + 5.2^\circ\ (c\ 0.745\ CHCl_3)\); lit. \([7]\), \([\alpha]_D^{25} + 5.1^\circ\ (c\ 3.51,\ CHCl_3)\), identical with that already prepared by Momose. The preparation of carbamate 14 constitute \([8]\) a formal synthesis of (-)-gephyrotoxin-223 [(−)-indolizidine \(223\text{AB}\)] (scheme 5).

**EXPERIMENTAL**

IR spectra were recorded using a Bomen MB-100 spectrophotometer. NMR spectra were recorded in CDCl\(_3\) solutions at 300 MHz (\(\text{H}\)), 282 MHz (\(^{19}\text{F}\)), 75 MHz (\(^{13}\text{C}\)) on a Bruker AC–300 instrument. Optical rotation
Scheme 3: Enzyme-catalyzed acetylation of cis-diol 3 with CAL.

Scheme 4: Chemoenzymatic enantioselective formal synthesis of (-)-gephyrotoxin-223.
values were obtained from a JASCO DIP-300 polarimeter (c as g of compound per 100 mL). Elemental analyses performed on a Carlo Ebra 1106 instrument. Column purifications were conducted by flash chromatography on silica gel 60 (230-400 mesh).

Enzyme-catalyzed acylation of N-carbobenzoxy-cis-2,6-dihydroxymethyl-piperidine 3: preparation of N-carbobenzoxy-cis-2S-acetoxymethyl-6R-hydroxymethyl-piperidine 1

To a solution of diol 3 (53 mg, 0.19 mmol) in vinylacetate (3 mL) was added 60 mg of Candida antarctica lipase (CAL), and the mixture was stirred for 3 h at rt. The progress of the reaction was monitored by TLC. As the reaction proceeded, the amount of diacetate present in the reaction mixture increased before the complete disappearance of starting material was observed. When the spot of diacetate became as visible as the spot of the starting diol, the reaction was stopped. The mixture was filtered for removing of enzyme and then concentrated. Purification of the crude product was done by flash chromatography using 20 % ethyl acetate / 80 % petroleum ether to pure ethyl acetate to give (2S,6R)-monoacetate 1 as a colourless oil in 80 % yield (48.81 mg, 0.151 mmol) with an ee ≥ 95 % as measured by 19F NMR of R-Mosher's ester made from 1. [α]D25 - 4.98 (c 2.08, CHCl3); IR (neat) vmax: 3450 (OH), 2930, 1745 (C=O, Ac), 1690 (C=O, NCO2) cm⁻1; 1H NMR (CDCl3) δ: 7.35 - 7.25 (m, 5H, Ph), 5.12 (AB system, 2H, J = 12.4 Hz, CH2Ph), 4.48 (m, 1H, CH2), 4.31 (m, 1H, CH2), 4.13 (dd, 1H, J1 = 8.0 Hz, J2 = 10.9 Hz, CHHOAc), 3.95 (dd, 1H, J1 = 6.9 Hz, J2 = 10.9 Hz, CHHOAc), 3.56 (d, 2H, J = 7.6 Hz, CH2OH), 2.85 (br s, 1H, OH), 1.92 (s, 3H, CH3CO), 1.81-1.45 (m, 6H, C3H2, C4H2 and C5H2); 13C
Preparation of N-carbobenzoxy- cis- 2R- (trans-carboethoxyethylene)-6S-(1-propenyl)-piperidine 12

The synthesis of alcohol 10 was reported from chiral synthon 1 in our previous article [3] and then it will transformed to the aldehyde 11 as following experiment. Oxalyl chloride (2 eq, 4 mmol, 0.34 mL) was dissolved in 6 mL anhydrous CH2Cl2, cooled to -78°C and stirred under N2. To this solution, anhydrous DMSO (3 eq, 6 mmol, 0.43 mL) in 2 mL anhydrous CH2Cl2 was added dropwise during 10 min and stirred to react for 5 min at -78 °C. The alcohol 10 (2 mmol, 438 mg) in 2 mL anhydrous CH2Cl2 was added dropwise to the reaction mixture, which was stirred for 1 hr at -78°C. The reaction was completed by addition of anhydrous Et3N (4 eq, 1.12 mL, 8 mmol). After 5 min, the dry ice/acetone bath was removed and the reaction temperature was left to rise to rt. The reaction was diluted with 10 mL of CH2Cl2 and then poured into 30 mL CH2Cl2 / 10 mL 10 % NH4OH solution. The aqueous phase was extracted 3 times with CH2Cl2, and the combined CH2Cl2 fractions were washed with brine and dried with MgSO4. Evaporation of the solvent gave a mixture of a light yellow oil and a white solid. After dissolving the oily product in anhydrous ether, the mixture was filtered through a MgSO4 pad and then the ether evaporated to give the highly unstable aldehyde 11, which was used immediately in the Wittig reaction. The purification and analysis of the resulting aldehyde 11 was not possible because it decomposes very fast.

Wittig reaction of aldehyde 11

To a solution of crude aldehyde 11 (2.0 mmol, assuming 100 % yield from Swern oxidation of alcohol 10) in anhydrous benzene (40 mL), ethyl (triphenylphosphoranylidene) acetate (1.5 eq, 3.0 mmol) was added. The reaction mixture was refluxed for 4 hr, and then poured into the solution of 50 mL ethyl acetate / 25 mL 10 % Na2SO3. The aqueous phase was extracted 3 times with ethyl acetate. The organic fractions were dried with MgSO4 and evaporated. The residue was purified by flash chromatography using 5% ethyl acetate / 95 % petroleum ether to 15 % ethyl acetate / 85 % petroleum ether to give 12 as a white solid in 80 % yield (437.4 mg, 1.6 mmol) from alcohol 10. [α]D^25 -105.9 (c 0.375, CHCl3) (from CAL); IR (neat) ν max : 3050 - 2944, 1717 (CO, CO2Et), 1693 (CO, NCO2), 1655 (C=C) cm^-1; 1H NMR (CDCl3) δ: 7.95 - 7.23 (m, 5H, Ph), 6.97 – 6.90 (dd, 1H, J1 = 5.1 Hz, J2 = 16 Hz, C10H), 5.89 (dd, 1H, J1 = 2 Hz, J2 = 16 Hz, C11H); 5.57 – 5.35 (m, 2H, C7H and C8H), 5.11 – 5.02 (m, 3H, CH2Ph and C2H), 4.90 (m, 1H, C6H), 4.16 – 4.09 (q, J = 7.1 Hz, CH2, Et), 1.89 – 1.47 (m, 6H, C3H2, C4H2 and C5H2), 1.55 (d, 3H, J= 6.42 Hz, C9H3), 1.25 – 1.20 (t, J= 7.23 Hz, CH3, Et). 13C NMR (CDCl3) δ: 166.29 (C=O, CO2Et), 155.59 (C=O, NCO2), 149.04 (C10), 136.45 (C5), 130.16 (C7), 128.31 (C9), 127.92 (C1), 127.87 (C2), 125.70 (C8), 121.43 (C11), 67.29 (CH3Ph), 60.28 (OCH2CH3), 50.77 (C5), 47.59 (C6), 30.04 (C3), 27.58 (C3), 14.96 (C4), 14.12 (CH3), 12.71 (CH3); HRMS (CI, NH3) calcd. (MH+) : 358.2018, Found (MH+): 358.2025 ±0.0011.

NMR (CDCl3) δ: 170.60 (C=O, Ac), 156.79 (C=O, NCO2), 136.43 (C1′), 128.36 (C3′, C5′), 127.89 (C4′), 127.72 (C2′, C6′), 67.28 (CH2OAc), 64.38 (CH3Ph), 64.02 (CH3OH), 51.71 (C6), 48.45 (C5), 24.84 (C3), 24.32 (C5), 20.5 (C4), 14.47 (CH3).
Preparation of (2R,6R)-N-carbobenzoxy-cis-2-(carboethoxyethyl)-6-propylpiperidine 14

Hydrogenation of diene 12

A suspension of diene 12 (50 mg, 0.14 mmol) and palladium hydroxide (10 mg) in EtOH (5 mL) was stirred under a hydrogen atmosphere overnight at 40 psi. After filtration of the reaction mixture through a bed of Celite, the filtrate was evaporated to give crude amino ester 13. It was used without further purification in the next step.

N-Protection of amino-ester 13

To a solution of crude amino-ester 13 in anhydrous THF (2 mL) under dry N2 at 0°C was added disopropylethylamine (DiPEA) (1.3 eq, 38.2 µL) followed by benzylchloroformate (1.2 eq, 30 µL). The ice bath was immediately removed and the reaction was stirred for 4 hr at room temperature. The mixture was diluted by addition CH2Cl2 (10 mL). To this solution was added 1N HCl (2 mL) and then extraction was effected 3 times with CH2Cl2. The combined organic fractions were dried with MgSO4 and evaporated. The crude product of 14 was purified by flash chromatography using 5 % ethyl acetate / 95 % hexane to 30 % ethyl acetate / 70 % hexane to provide 14 (48.07 mg, 0.133 mmol) as an oil in 95 % yield. [α]D25: -5.2 (c 0.745, CHCl3) (from CAL), lit.[7] [α]D25: 5.1° (c 3.51, CHCl3); IR (neat) νmax: 2944, 1718 (CO, CO2Et), 1692 (CO, NCO2) cm-1; 1H NMR (CDCl3) δ: 7.35 - 7.27 (m, 5H, Ph), 5.17 – 5.04 (AB system, 2H, J = 12.42 Hz, CH2Ph), 4.21 (m, 2H, C3H, C6H), 4.11 – 4.07 (m, 2H, CH2, Et), 2.29 (m, 2H, C11H2), 1.95 – 1.79 (m, 2H, C10H2), 1.60 – 1.40 (m, 8H, C3H2, C4H2, C5H2, and C7H2), 1.24 - 1.13 (m, 2H, C8H2), 1.24 – 1.20 (t, 3H, J = 6.4 Hz, C13H3, Et), 0.90 – 0.86 (t, 3H, J = 7 Hz, C9H3). 13C NMR (CDCl3) δ: 173.2 (C=O, CO2), 155.98 (C=O, NCO2), 137.04 (C1′), 128.29 (C3′, C5′), 127.70 (C2′, C4′, C6′), 66.86 (CH2Ph), 60.19 (C5), 50.46 (C6), 49.94 (C3), 36.79 (C7), 32.03 (C5′), 29.70 (C3′), 29.56 (C10′), 28.05 (C11′), 27.36 (C3′), 14.19 (C8), 14.07 (C13H3), 13.86 (C1′)

CONCLUSION

In this work, we wished to show the effectiveness of enzymes in the enantioselective synthesis of (-)-gephyrotoxin-223. At first, meso and cis-2,6-piperidines 2, 3, were prepared from commercially available pyridine 2,6-dicarboxylic acid 4. The enzymatic hydrolysis of N-carbobenzoxy-cis-2,6-diacetoxymethyl-piperidine 2 with Aspergillus niger lipase gave (2R,6S)-monoacetate 1, N-carbobenzoxy-cis-2R-acetoxymethyl-6S-hydroxymethylpiperidine, in 5 days with good yield (83 %) and excellent enantiomeric excess (ee ≥ 98 %). The enzymatic acetylation of meso-diol 3, N-carbobenzoxy-cis-2,6-dihydroxy-methylpiperidine employing vinyl acetate as solvent and acyl donor in the presence of Candida antarctica lipase gave (2S,6R)-monoacetate 1, N-carbobenzoxy-cis-2S-acetoxymethyl-6R-hydroxymethylpiperidine, in only 3 h in good yield (80 %) and high enantiomeric purity (ee ≥ 95 %). In this manner, the time of desymmetrization has been decreased from 5 days to just 3 hours, which was an excellent improvement. By exploiting these two desymmetrization methods, we have synthesized the two enantiomeric pairs of chiral synthon 1, which were used in the synthesis of both enantiomeric pairs of several piperidines alkaloids (scheme 5). This is an eloquent demonstration of the beauty of our work.

Thus, in this work, the both enantiopure (-)-gephyrotoxin-223 were formally synthesized from pyridine-2,6-dicarboxylic acid 4.

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